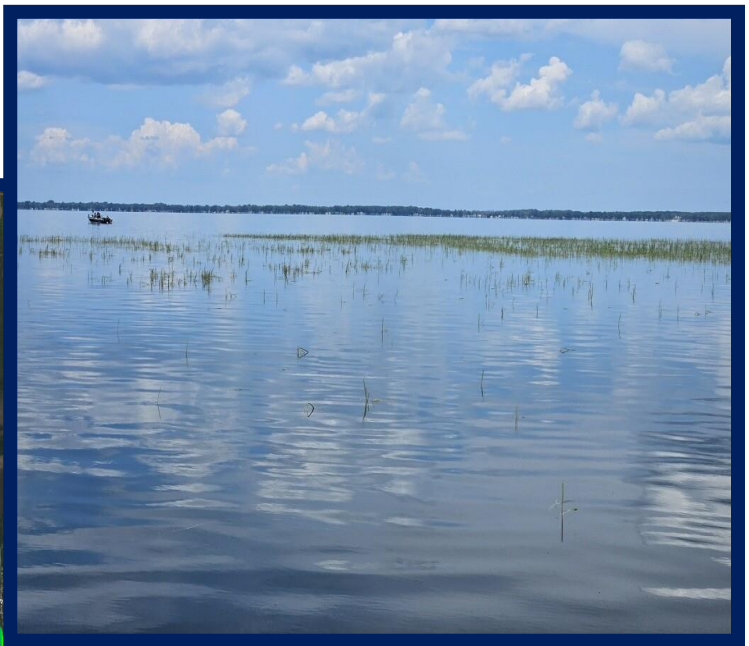
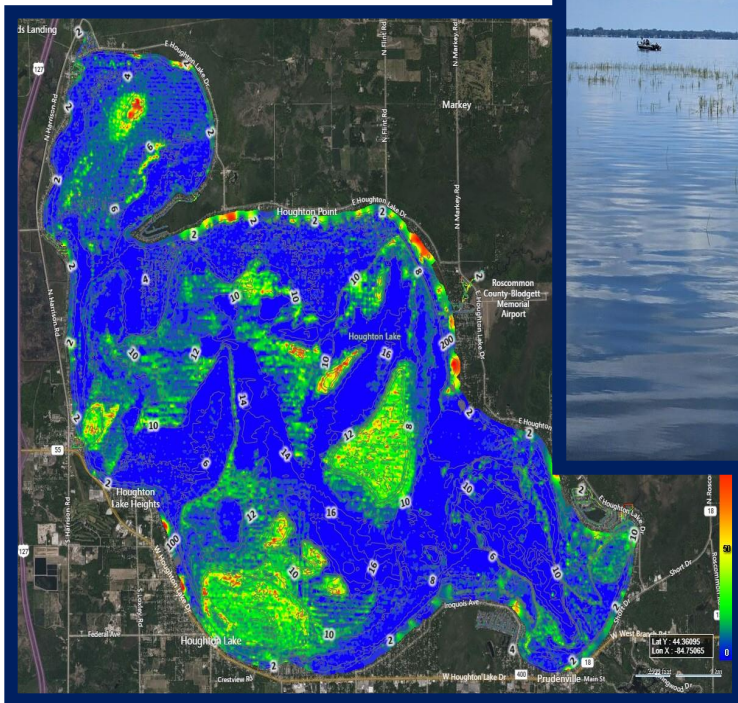




Houghton Lake 2024 Annual Report with 2025 Management Recommendations



February, 2025

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Houghton Lake 2024 Annual Report with 2025 Management Recommendations



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18406 W. Spring Lake Road
Spring Lake, Michigan 49456
Website: <http://www.restorativelakesciences.com>**

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Houghton Lake 2024 Annual Report with 2025 Management Recommendations

The following Houghton Lake report is a summary of key lake findings collected in 2024 with recommendations for continued improvement

The overall condition of Houghton Lake has been improving over the past few years due to rigorous aquatic vegetation surveys and selective spot-treatments to control invasive aquatic plant species such as hybrid Eurasian Watermilfoil (EWM) and Starry Stonewort. Both of these species are declining in Houghton Lake and providing space for the now 37 native aquatic plant species that are so important to the ecological balance of Houghton Lake. This represents an increase in 6 species relative to 2023! A total of 332 acres of lake area were treated for invasive aquatic plants in 2024. There was significant natural germination and colonization of 40 acres of Wild Rice in the Middle Grounds area which was possible due to effective sustainable control of the milfoil in that region. Additionally, significant growth of another favorable native, Water Marigold was also prevalent in the Middle Grounds due to reduction of the milfoil over the years. Continued growth of the Wild Rice in this region will allow for reduced colonization of milfoil or other invasives over time. If the natural population declines, seeding may be necessary and recommended along with the area in the North Bay.

The water quality of Houghton Lake in 2024 was excellent. The mean total phosphorus concentration in September of 2024 was 0.010 mg L^{-1} which is quite low and favorable. The mean total Kjeldahl nitrogen concentration was 0.6 mg L^{-1} which is also low and favorable. Nutrient concentrations in the lake were much lower in 2024 due to less runoff from reduced rainfall events. This indicates that the Houghton Lake ecosystem may be sensitive to runoff as a primary source of nutrients, especially from storm drains. The canals and tributaries have significantly higher nutrients than the lake basin and are thus a source.

There was abundant sunlight in 2024, and this has resulted in a favorable increase in submersed native aquatic vegetation which was one of the original feasibility improvement goals. Secchi transparency of water clarity were also higher in 2024 with a mean of 7.5 feet. Other parameters such as dissolved oxygen, pH, and conductivity were ideal for an inland lake. The algal communities were diverse and a good source of primary productivity for the fishery.

The presence of blue-green algae (*Microcystis* sp.) in some of the canals (McKinley and Long Point) is concerning and all canals will be monitored for these blooms and treated if necessary. RLS also determined the presence of the green filamentous algae, *Cladophora* which was found in a few locations nearshore on rocks and concrete seawalls in 2023-2024. The populations of both algal types are prevalent in lake systems with abundant populations of Zebra Mussels. The sediment macroinvertebrate community was also healthy relative to taxa and relative abundance, and this may change annually due to environmental conditions. Given the ability of *Cladophora* to become a nuisance by fouling shoreline habitat and docks and hoists, RLS recommends a comprehensive baseline evaluation of *Cladophora* along the shoreline of Houghton Lake as well as additional measurements to possibly determine the likely causes of *Cladophora* in specific areas. RLS proposes to conduct this evaluation during the summer of 2025 and will issue a separate report of the findings and any needed mitigation recommendations.

RLS recommends continued intense aquatic vegetation community surveys of the entire lake and canals and spot-treatments as needed for management of invasive species only. The canals and Middle Grounds areas have proven to require earlier survey dates than the remainder of the lake due to germination patterns. Rotation of herbicides is important to reduce the probability of tolerance to one used in a given year for a specific area. Surprisingly, Wild Rice colonization was naturally strong in the Middle Grounds in 2023-2024 and detailed surveys will be conducted again in 2025 to evaluate additional colonization and re-growth. There were approximately 40 acres of Wild Rice in the Middle Grounds located in 2024 which is a sign of strong natural colonization that may be possible due to lower water levels and also sustained control of the once-prevalent EWM in that region. The North Bay is also showing some natural growth and this area would be a suitable candidate for future seedlings if sources are available. There are some additional areas of strong natural growth in the Muskegon River that were documented in 2024 and will also be evaluated. Wild Rice was planted in Muddy Bay in 2020 and showed a 98.8% germination success in 2021.

However, by the end of 2023 due to a lack of available seed source, the population declined to 8.8% with a slight rebound to 11.3% in 2024. These areas will be monitored for long-term changes in the Wild Rice communities.

The Muddy Bay location may not be the ideal location due to the influences of strong fetch and ice scouring that limit germination. RLS determined that the Wild Rice locations had significantly higher nutrient concentrations than other areas in the main Basin and this explains why those populations prefer those locations for sustained growth. Future seeding, if conducted should be aligned with the statewide Wild Rice Stewardship Initiative program with local strains. There are current efforts statewide to enhance the growth and distribution of Wild Rice in state waters.

Key restoration and management recommendations were provided in the 2022 feasibility report and remain consistent for 2024. Pending data collection on the lake in 2024 will determine if those recommendations need any modifications. A community-wide workshop is planned for 2025 that will provide education and demonstrations for lakefront owners and lake users.

Houghton Lake Water Quality Data (2024)

Water Quality Parameters Measured

There are numerous water quality parameters that can be measured on an inland lake, but several are the most critical indicators of lake health. The parameters measured in Houghton Lake in 2024 and in previous years included: water temperature (measured in °C or °F), dissolved oxygen (measured in mg/L), pH (measured in standard units-SU), conductivity (measured in micro-Siemens per centimeter- $\mu\text{S}/\text{cm}$), total alkalinity or hardness (measured in mg of calcium carbonate per liter-mg CaCO_3/L), total dissolved solids (mg/L), secchi transparency (feet), total phosphorus, ortho-phosphorus, and total Kjeldahl nitrogen (all in mg/L), chlorophyll-a (in $\mu\text{g}/\text{L}$), and algal community composition. Graphs that show trends for some parameters of each year are displayed below. Water quality was measured in the deep basins of Houghton Lake on September 18, 2024 (Figure 1). Additional water quality samples were collected in the tributaries (Figure 2) and in the canals (Figure 3).

Trend data was calculated using mean values of the most critical parameters over the sampling locations and mean values are represented as red lines on each graph (Figures 4-7). Table 1 below demonstrates how lakes are classified based on key parameters. Houghton Lake would be historically considered meso-eutrophic (relatively productive) since it does contain ample phosphorus, nitrogen, and aquatic vegetation growth but has good water clarity and moderately low planktonic algal growth. General water quality classification criteria are defined in Table 1. 2024 water quality data for Houghton Lake are shown below in Tables 2-7. Water quality data for the tributaries and canals are shown in Tables 8-12.

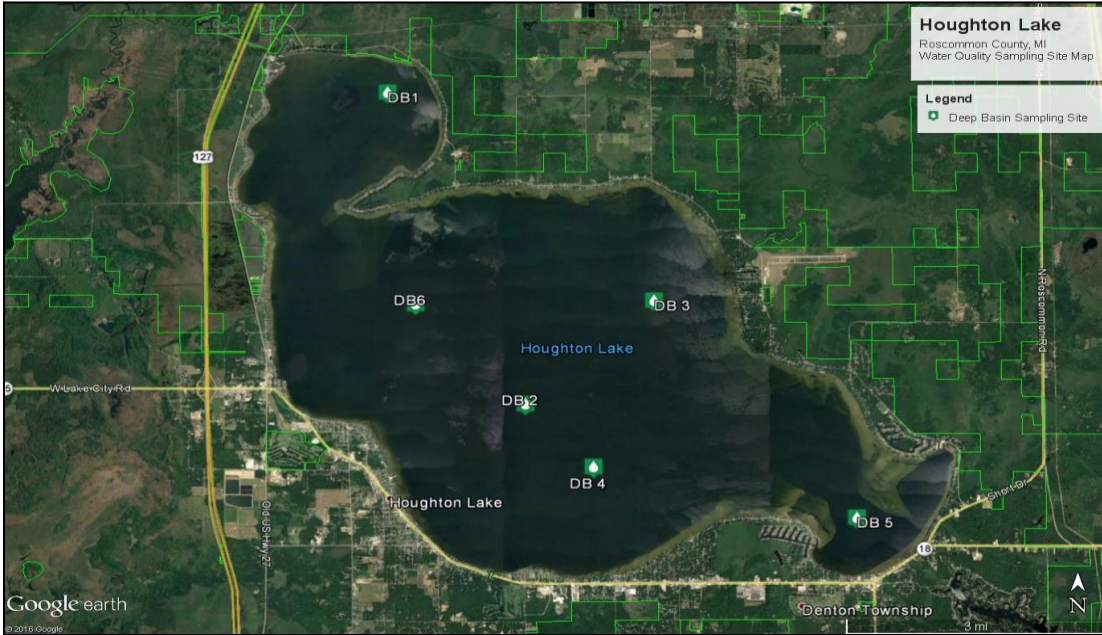


Figure 1. Deep basin water quality sampling locations in Houghton Lake (2016-2024).

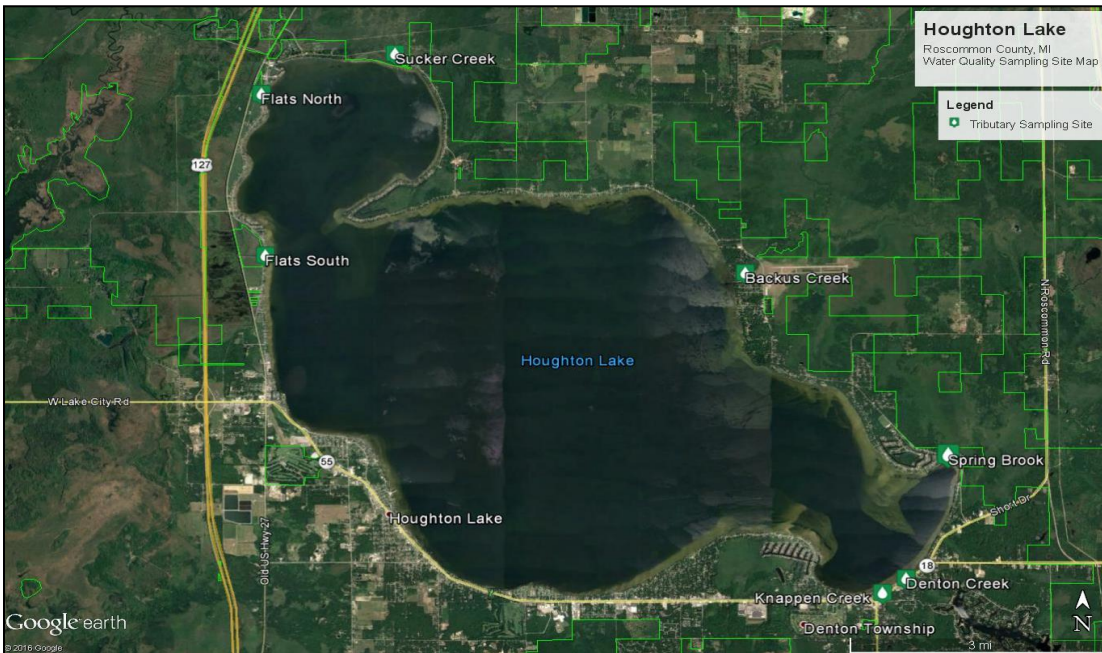


Figure 2. Tributary water quality sampling locations around Houghton Lake (2016-2024).

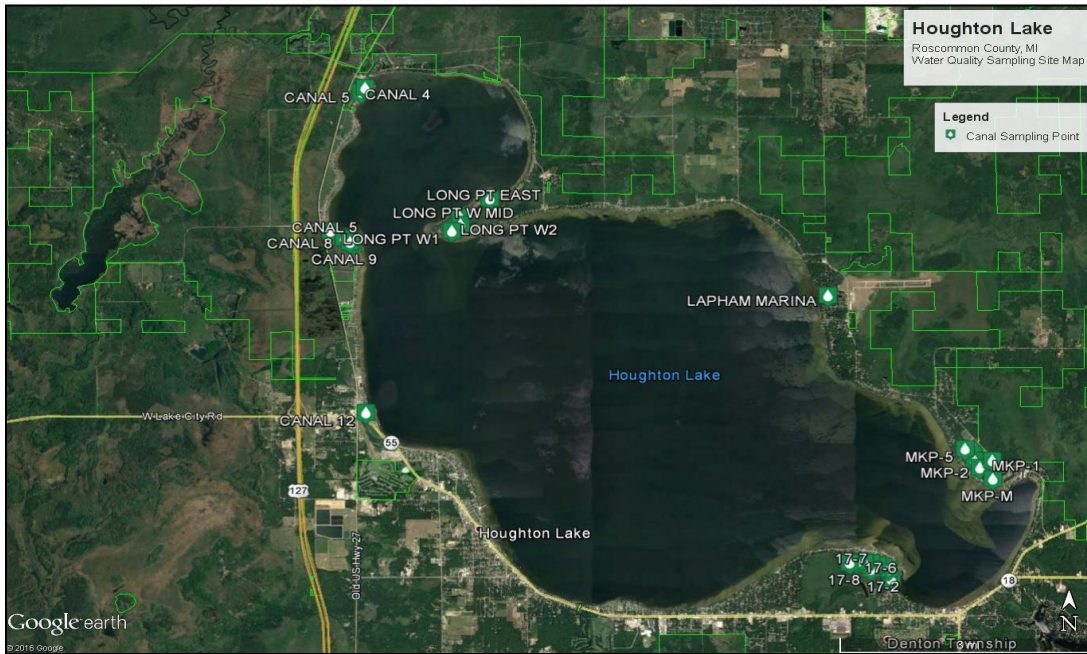


Figure 3. Houghton Lake canals water quality sampling locations (2016-2024).

Table 1. Lake trophic classification (MDNR).

<i>Lake Trophic Status</i>	<i>Total Phosphorus ($\mu\text{g L}^{-1}$)</i>	<i>Chlorophyll-a ($\mu\text{g L}^{-1}$)</i>	<i>Secchi Transparency (feet)</i>
Oligotrophic	< 10.0	< 2.2	> 15.0
Mesotrophic	10.0 – 20.0	2.2 – 6.0	7.5 – 15.0
Eutrophic	> 20.0	> 6.0	< 7.5

Houghton Lake Deep Basin Water Quality Data Tables:

Table 2. Houghton Lake water quality parameter data collected in deep basin #1 on September 18, 2024.

Depth ft.	Water Temp °C	DO mg L ⁻¹	pH S.U.	Cond μS cm ⁻¹	Turb NTU	TDS mg L ⁻¹	TP mg L ⁻¹	Ortho-P mg L ⁻¹	TKN mg L ⁻¹	Chl-a μg L ⁻¹	Secchi (ft)
0	22.5	8.7	8.3	238	0.4	153	<0.010	<0.010	0.7	--	--
2.5	22.0	9.3	8.5	235	0.7	151	<0.010	<0.010	0.7	2.0	5.4+
5.0	22.0	9.9	8.5	236	0.9	150	<0.010	<0.010	0.7	--	--

Table 3. Houghton Lake water quality parameter data collected in deep basin #2 on September 18, 2024.

Depth ft.	Water Temp °C	DO mg L ⁻¹	pH S.U.	Cond μS cm ⁻¹	Turb NTU	TDS mg L ⁻¹	TP mg L ⁻¹	Ortho-P mg L ⁻¹	TKN mg L ⁻¹	Chl-a μg L ⁻¹	Secchi (ft)
0	21.1	9.1	8.7	243	0.3	155	<0.010	<0.010	0.5	--	--
8.0	21.0	9.2	8.7	243	0.8	155	<0.010	<0.010	0.5	2.0	8.3
16.0	20.7	9.2	8.7	243	0.9	155	<0.010	<0.010	0.5	--	--

Table 4. Houghton Lake water quality parameter data collected in deep basin #3 on September 18, 2024.

Depth ft.	Water Temp °C	DO mg L ⁻¹	pH S.U.	Cond μS cm ⁻¹	Turb NTU	TDS mg L ⁻¹	TP mg L ⁻¹	Ortho-P mg L ⁻¹	TKN mg L ⁻¹	Chl-a μg L ⁻¹	Secchi (ft)
0	21.1	9.0	8.6	247	0.3	158	<0.010	<0.010	0.6	--	--
8.0	20.9	9.1	8.6	247	0.4	158	<0.010	<0.010	0.5	3.0	8.5
16.0	19.3	6.8	7.9	287	0.7	157	<0.010	<0.010	0.6	--	--

Table 5. Houghton Lake water quality parameter data collected in deep basin #4 on September 18, 2024.

<i>Depth ft.</i>	<i>Water Temp °C</i>	<i>DO mg L⁻¹</i>	<i>pH S.U.</i>	<i>Cond μS cm⁻¹</i>	<i>Turb NTU</i>	<i>TDS mg L⁻¹</i>	<i>TP mg L⁻¹</i>	<i>Ortho-P mg L⁻¹</i>	<i>TKN mg L⁻¹</i>	<i>Chl-a μg L⁻¹</i>	<i>Secchi (ft)</i>
0	20.8	9.0	8.5	242	0.2	155	<0.010	<0.010	0.7	--	--
10.0	20.8	9.1	8.5	242	0.5	155	<0.010	<0.010	0.8	2.0	8.0
20.0	20.1	9.3	8.6	244	0.6	156	<0.010	<0.010	0.5	--	--

Table 6. Houghton Lake water quality parameter data collected in deep basin #5 on September 18, 2024.

<i>Depth ft.</i>	<i>Water Temp °C</i>	<i>DO mg L⁻¹</i>	<i>pH S.U.</i>	<i>Cond μS cm⁻¹</i>	<i>Turb NTU</i>	<i>TDS mg L⁻¹</i>	<i>TP mg L⁻¹</i>	<i>Ortho-P mg L⁻¹</i>	<i>TKN mg L⁻¹</i>	<i>Chl-a μg L⁻¹</i>	<i>Secchi (ft)</i>
0	20.4	9.2	8.6	245	0.3	157	<0.010	<0.010	0.6	--	--
10.0	19.9	9.3	8.6	246	0.7	157	<0.010	<0.010	0.6	2.0	6.5
20.0	18.2	6.7	8.0	260	0.9	168	0.016	<0.010	0.6	--	--

Table 7. Houghton Lake water quality parameter data collected in deep basin #6 on September 18, 2024.

<i>Depth ft.</i>	<i>Water Temp °C</i>	<i>DO mg L⁻¹</i>	<i>pH S.U.</i>	<i>Cond μS cm⁻¹</i>	<i>Turb NTU</i>	<i>TDS mg L⁻¹</i>	<i>TP mg L⁻¹</i>	<i>Ortho-P mg L⁻¹</i>	<i>TKN mg L⁻¹</i>	<i>Chl-a μg L⁻¹</i>	<i>Secchi (ft)</i>
0	21.5	9.3	8.2	240	0.3	153	<0.010	<0.010	0.6	--	--
6.0	21.4	9.4	8.2	239	0.7	153	0.010	<0.010	0.6	2.0	8.5
12.0	21.3	9.5	8.5	239	0.9	153	0.010	<0.010	0.5	--	--

Houghton Lake Canal Water Quality Data Tables:

Table 8. Houghton Lake water quality parameter data collected in the Chippewa canals on September 18, 2024. Note: All samples were collected at a mid-depth of 3.0 feet. Site CM refers to the middle of the canal series.

<i>Canal Site</i>	<i>Water Temp °C</i>	<i>DO mg L⁻¹</i>	<i>pH S.U.</i>	<i>Cond. µS cm⁻¹</i>	<i>Turb. NTU</i>	<i>TDS mg L⁻¹</i>	<i>TP mg L⁻¹</i>	<i>Chl-a µg L⁻¹</i>	<i>Secchi (ft)</i>
C1	19.5	5.8	7.8	559	3.1	358	0.010	7.0	4.7
C2	19.1	4.7	7.8	598	3.5	386	0.026	8.0	6.2
C3	18.9	4.4	7.8	602	3.0	385	0.026	7.0	5.8
C4	20.2	7.3	8.1	577	2.5	369	0.036	7.0	3.0
C5	21.2	9.4	8.4	573	4.7	366	0.036	10.0	2.8
C6	21.3	7.8	8.5	574	3.2	367	0.034	9.0	2.8
C7	19.0	7.5	8.0	599	4.9	283	0.034	9.0	2.3
C8	19.7	6.5	8.1	591	3.2	378	0.038	9.0	3.1
CM	21.6	8.5	8.4	569	3.4	364	0.036	11.0	2.7

Table 9. Houghton Lake water quality parameter data collected in the McKinley Park (MPK) canals on September 18, 2024. Note: All samples were collected at mid-depth of 3.0 feet. Site MPK M refers to the middle of the canal series.

<i>Canal Site</i>	<i>Water Temp °C</i>	<i>DO mg L⁻¹</i>	<i>pH S.U.</i>	<i>Cond. µS cm⁻¹</i>	<i>Turb. NTU</i>	<i>TDS mg L⁻¹</i>	<i>TP mg L⁻¹</i>	<i>Chl-a µg L⁻¹</i>	<i>Secchi (ft)</i>
MPK 1	20.7	6.7	7.9	339	2.8	217	0.026	4.0	4.0
MPK 2	20.6	7.5	8.0	355	4.2	227	0.016	10.0	6.2
MPK 3	20.5	2.8	7.6	380	4.6	243	0.024	8.0	5.3
MPK 4	19.6	3.9	7.6	387	4.2	248	0.010	7.0	5.6
MPK 5	20.7	3.6	7.6	381	4.1	244	0.022	7.0	3.0
MPK M	21.4	6.0	7.9	349	2.2	223	0.010	10.0	5.4

Table 10. Houghton Lake water quality parameter data collected in the Lapham and Long Point canals on September 18, 2024. Note: All samples were collected at mid-depth of 3.0 feet.

<i>Canal Site</i>	<i>Water Temp °C</i>	<i>DO mg L⁻¹</i>	<i>pH S.U.</i>	<i>Cond. µS cm⁻¹</i>	<i>Turb. NTU</i>	<i>TDS mg L⁻¹</i>	<i>TP mg L⁻¹</i>	<i>Chl-a µg L⁻¹</i>	<i>Secchi (ft)</i>
LAPHAM	21.6	8.5	8.3	557	1.9	357	0.036	14.0	3.7
L POINT MID	20.2	8.2	8.0	282	3.2	186	0.024	12.0	6.3
L POINT W1	19.5	4.7	7.2	366	2.3	235	0.046	14.0	5.8
L POINT W2	18.6	6.3	7.7	360	2.0	230	0.030	10.0	5.8
L POINT E	17.0	5.1	7.2	366	1.9	234	0.042	9.0	4.9

Table 11. Houghton Lake water quality parameter data collected in the canals north and west of Long Point canals #4-12 on September 18, 2024. Note: All samples were collected at mid-depth of 3.0 feet.

<i>Canal Site</i>	<i>Canal Name</i>	<i>Water Temp °C</i>	<i>DO mg L⁻¹</i>	<i>pH S.U.</i>	<i>Cond. µS cm⁻¹</i>	<i>Turb. NTU</i>	<i>TDS mg L⁻¹</i>	<i>TP mg L⁻¹</i>	<i>Chl-a µg L⁻¹</i>	<i>Secchi (ft)</i>
CANAL 4	Porath	22.2	5.2	7.3	299	1.6	191	0.180	5.0	2.5
CANAL 5	Ford	22.3	8.1	8.2	263	1.9	168	0.046	8.0	5.0
CANAL 6	Holt	22.7	5.0	7.4	299	1.7	192	0.042	8.0	5.0
CANAL 8	Siebert	21.7	7.1	7.8	266	1.9	170	0.030	3.0	4.4
CANAL 9	Church	22.8	7.4	8.0	264	2.6	169	0.018	4.0	3.9
CANAL 10	Swick	22.6	6.7	7.7	280	2.3	180	0.030	4.0	3.9
CANAL 12	Beebe	22.8	5.8	7.6	267	2.7	172	0.010	3.0	2.7

Houghton Lake Tributary Water Quality Data Table:

Table 12. Houghton Lake water quality parameter data collected in the tributaries and flats on September 18, 2024.

<i>Tributary Site</i>	<i>Water Temp °C</i>	<i>DO mg L⁻¹</i>	<i>pH S.U.</i>	<i>Cond. µS cm⁻¹</i>	<i>Turb. NTU</i>	<i>TDS mg L⁻¹</i>	<i>TSS mg L⁻¹</i>	<i>TP mg L⁻¹</i>	<i>TKN mg L⁻¹</i>
DENTON CREEK	--	--	--	--	--	--	--	--	--
SPRING BROOK	--	--	--	--	--	--	--	--	--
BACKUS/CUT	21.2	8.4	8.3	280	1.9	179	<10	<0.010	<0.5
KNAPPEN CREEK	22.9	8.5	8.1	262	2.3	168	<10	<0.010	<0.5

Dissolved Oxygen

Dissolved oxygen is a measure of the amount of oxygen that exists in the water column. In general, dissolved oxygen levels should be greater than 5 mg L⁻¹ to sustain a healthy warm-water fishery. Dissolved oxygen concentrations may decline if there is a high biochemical oxygen demand (BOD) where organismal consumption of oxygen is high due to respiration. Dissolved oxygen is generally higher in colder waters. Dissolved oxygen was measured in milligrams per liter (mg L⁻¹) with the use of a calibrated Eureka Manta II® dissolved oxygen meter. During the summer months, dissolved oxygen at the surface is generally higher due to the exchange of oxygen from the atmosphere with the lake surface, whereas dissolved oxygen is lower at the lake bottom due to decreased contact with the atmosphere and increased biochemical oxygen demand (BOD) from microbial activity. Dissolved oxygen concentrations during the September 18, 2024 sampling event averaged 9.0 mg L⁻¹. Figure 4 below shows the changes in mean DO with time in Houghton Lake with a historic mean of 9.1 mg L⁻¹ which is excellent.

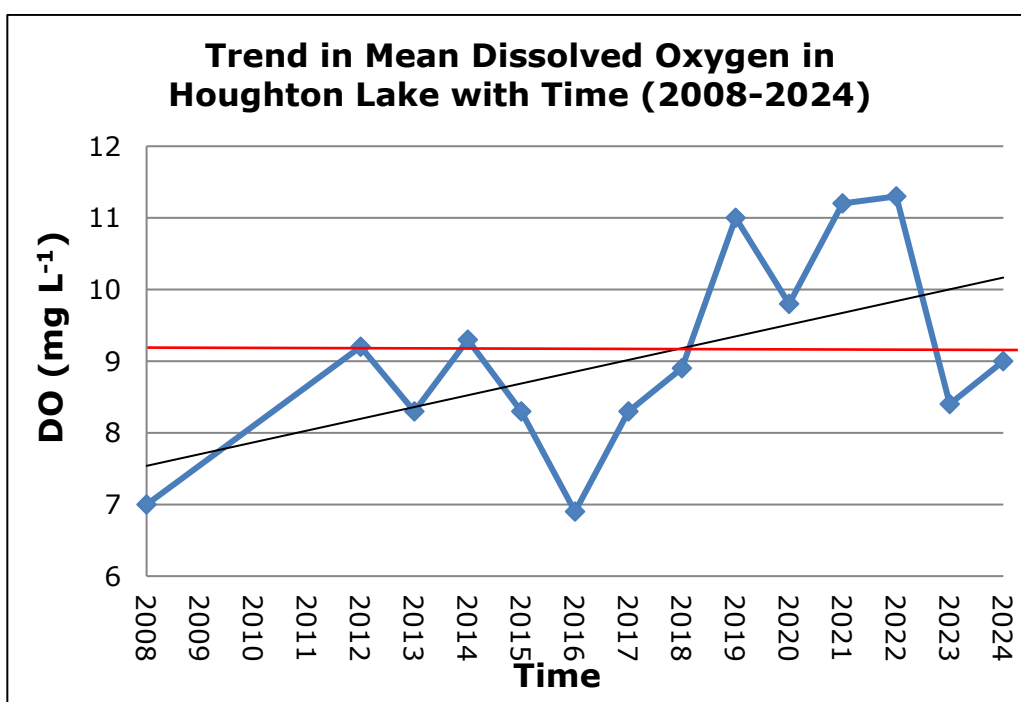


Figure 4. Changes in mean DO with time in Houghton Lake (2008-2024).

Water Clarity (Transparency)

Elevated Secchi transparency readings allow for more aquatic plant and algae growth. The transparency throughout Houghton Lake was adequate on September 18, 2024 (mean of 7.5 feet) to allow abundant growth of algae and aquatic plants in the majority of the littoral zone of the lake.

Secchi transparency is variable and depends on the number of suspended particles in the water (often due to windy conditions of lake water mixing) and the amount of sunlight present at the time of measurement. Other parameters such as turbidity (measured in NTU's) and Total Dissolved Solids (measured in mg/L) are correlated with water clarity and show an increase as clarity decreases. Figure 5 below displays the trend in mean Secchi transparency with time. The historic mean is 6.1 feet which is moderate but common with large, shallow, high-energy systems.

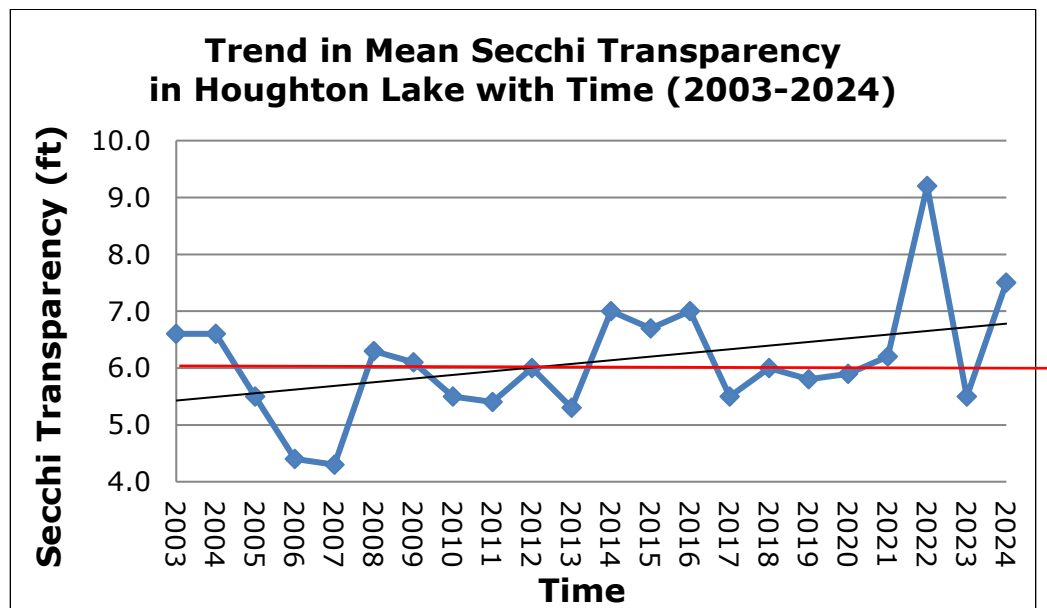


Figure 5. Changes in mean Secchi Transparency with time in Houghton Lake (2003-2024).

Total Phosphorus & Ortho-Phosphorus

Total phosphorus (TP) is a measure of the amount of phosphorus (P) present in the water column. Phosphorus is the primary nutrient necessary for abundant algae and aquatic plant growth. TP concentrations are usually higher at increased depths due to higher release rates of P from lake sediments under low oxygen (anoxic) conditions. Phosphorus may also be released from sediments as pH increases.

Fortunately, even though the TP levels in Houghton Lake are moderate, the dissolved oxygen levels are high enough at the bottom to not result in the release of phosphorus from the bottom. The mean TP concentration on September 18, 2024 was 0.010 mg L^{-1} (Figure 6), which is lower than in recent years and was very well below the eutrophic threshold. Ortho-phosphorus or “soluble reactive phosphorus” refers to the proportion of phosphorus that is bioavailable to aquatic life. Higher concentrations of ortho-phosphorus concentrations in the lake result in increased uptake of the nutrient by aquatic plants and algae. The ortho-phosphorus concentrations in the deep basins of Houghton Lake were all $\leq 0.010 \text{ mg L}^{-1}$, which were quite low and also favorable. The mean TP in the canals was higher at 0.034 mg L^{-1} . The mean TP in the tributaries was much lower at $<0.010 \text{ mg L}^{-1}$. The historic mean in the lake basin is 0.018 mg L^{-1} or $18.0 \mu\text{g L}^{-1}$ which is low and favorable.

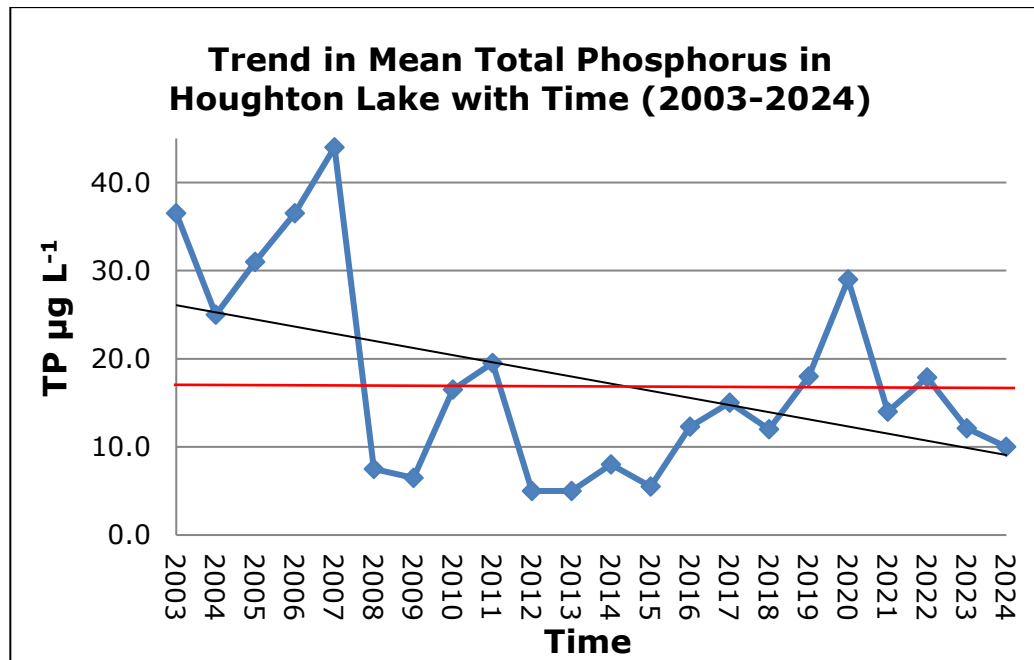


Figure 6. Changes in mean TP with time in Houghton Lake (2003-2024).

Total Kjeldahl Nitrogen

Total Kjeldahl Nitrogen (TKN) is the sum of ammonia (NH_3^+), and organic nitrogen forms in freshwater systems. Much nitrogen (amino acids and proteins) also comprises the bulk of living organisms in an aquatic ecosystem.

Nitrogen originates from atmospheric inputs (i.e., burning of fossil fuels), wastewater sources from developed areas (i.e., runoff from fertilized lawns), agricultural lands, septic systems, and from waterfowl droppings. It also enters lakes through ground or surface drainage, drainage from marshes and wetlands, or from precipitation (Wetzel, 2001). In lakes with an abundance of nitrogen ($N:P > 15$), phosphorus may be the limiting nutrient for phytoplankton and aquatic macrophyte growth. Alternatively, in lakes with low nitrogen concentrations (and relatively high phosphorus), the blue-green algae populations may increase due to the ability to fix nitrogen gas from atmospheric inputs. Lakes with a mean TKN value of 0.66 mg L^{-1} may be classified as oligotrophic, those with a mean TKN value of 0.75 mg L^{-1} may be classified as mesotrophic, and those with a mean TKN value greater than 1.88 mg L^{-1} may be classified as eutrophic. The mean TKN concentration in Houghton Lake on September 18, 2024 averaged 0.6 mg L^{-1} , which is moderately low for an inland lake and similar to last year. The historic mean for the lake basin is 0.7 mg L^{-1} , which is favorable (Figure 7). The TKN in the tributaries ranged from $<0.5 \text{ mg L}^{-1}$. Note that in 2024 at the time of sampling, only the Cut River and Knappen Creek were flowing due to low rainfall conditions. Tributaries are only sampled when they are flowing to allow for data representative of actively flowing conditions.

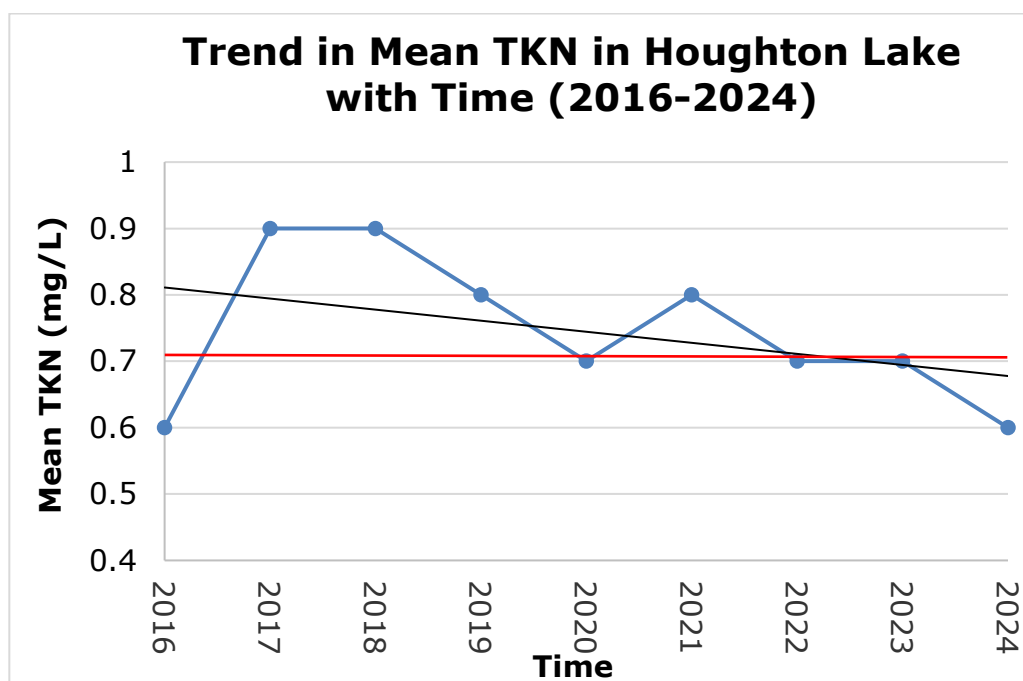


Figure 7. Changes in mean TKN with time in Houghton Lake (2016-2024).

Total Alkalinity

Lakes with high alkalinity ($>150 \text{ mg L}^{-1}$ of CaCO_3) are able to tolerate larger acid inputs with less change in water column pH. Many Michigan lakes contain high concentrations of CaCO_3 and are categorized as having “hard” water. Total alkalinity may change on a daily basis due to the re-suspension of sedimentary deposits in the water and respond to seasonal changes due to the cyclic turnover of the lake water. The alkalinity of Houghton Lake was moderately low on September 18, 2024 (mean of 86 mg L^{-1} of CaCO_3) and indicates a slightly soft-water lake. Total alkalinity is periodically evaluated to determine how the buffering capacity of the lake changes over time.

Turbidity & Total Dissolved Solids

Turbidity is a measure of the loss of water transparency due to the presence of suspended particles. The turbidity of water increases as the number of total suspended particles increases. Turbidity may be caused by erosion inputs, phytoplankton blooms, storm water discharge, urban runoff, re-suspension of bottom sediments, and in smaller lakes by large bottom-feeding fish such as carp. Particles suspended in the water column absorb heat from the sun and raise water temperatures. Since higher water temperatures generally hold less oxygen, shallow turbid waters are usually lower in dissolved oxygen. Turbidity was measured in Nephelometric Turbidity Units (NTU's) with the use of a calibrated turbidimeter. The World Health Organization (WHO) requires that drinking water be less than 5.0 NTU's; however, recreational waters may be significantly higher than that.

The turbidity of Houghton Lake was quite low and was ≤ 0.9 NTU's during the 2024 sampling event. Spring values may be higher due to increased watershed inputs from spring runoff and/or from increased algal blooms in the water column from resultant runoff contributions. The turbidity of the canals was ≤ 4.9 NTU's and was lower in 2024 due to less runoff associated nutrients and solids. The turbidity of the tributaries was ≤ 2.3 NTU's which is favorable.

Total dissolved solids (TDS) is a measure of the amount of dissolved organic and inorganic particles in the water column. Particles dissolved in the water column absorb heat from the sun and raise the water temperature and increase conductivity. TDS was measured with the use of a calibrated Eureka Manta II® TDS probe in mg L^{-1} . Spring values may be higher due to increased watershed inputs from spring runoff and/or increased planktonic algal communities. The TDS in Houghton Lake was $\leq 168 \text{ mg L}^{-1}$ for the deep basins on September 18, 2024, which is moderate for an inland lake.

The preferred range for TDS in surface waters is between 0-1,000 mg L⁻¹ but the lower values are most favorable. The TDS in the canals was ≤386 mg L⁻¹ which is higher than the lake and likely due to the presence of stormwater inputs and stagnant waters. The TDS of the tributaries was ≤179 mg L⁻¹.

Total Suspended Solids

Total suspended solids (TSS) refers to the quantity of solid particles detected in the water that reduce light penetration and create turbidity in the water. The TSS samples measured in the Houghton Lake tributaries were all ≤10 mg L⁻¹, which is low overall. The ideal concentration for TSS in inland lakes and streams is ≤20 mg L⁻¹. TSS may increase during periods of heavy rainfall/runoff.

pH

Most Michigan lakes have pH values that range from 6.5 to 9.5. Acidic lakes (pH < 7) are rare in Michigan and are most sensitive to inputs of acidic substances due to a low acid neutralizing capacity (ANC). Houghton Lake is considered “slightly basic” on the pH scale. The pH of Houghton Lake averaged 8.5 S.U. (Figure 8) on September 18, 2024, which is ideal for an inland lake. The pH of the canals ranged from 7.2-8.4 S.U. and the pH of the tributaries ranged from 8.1-8.3 S.U. All of these values are normal and favorable for aquatic environments. The historic mean pH in the lake basin is 8.4 S.U.

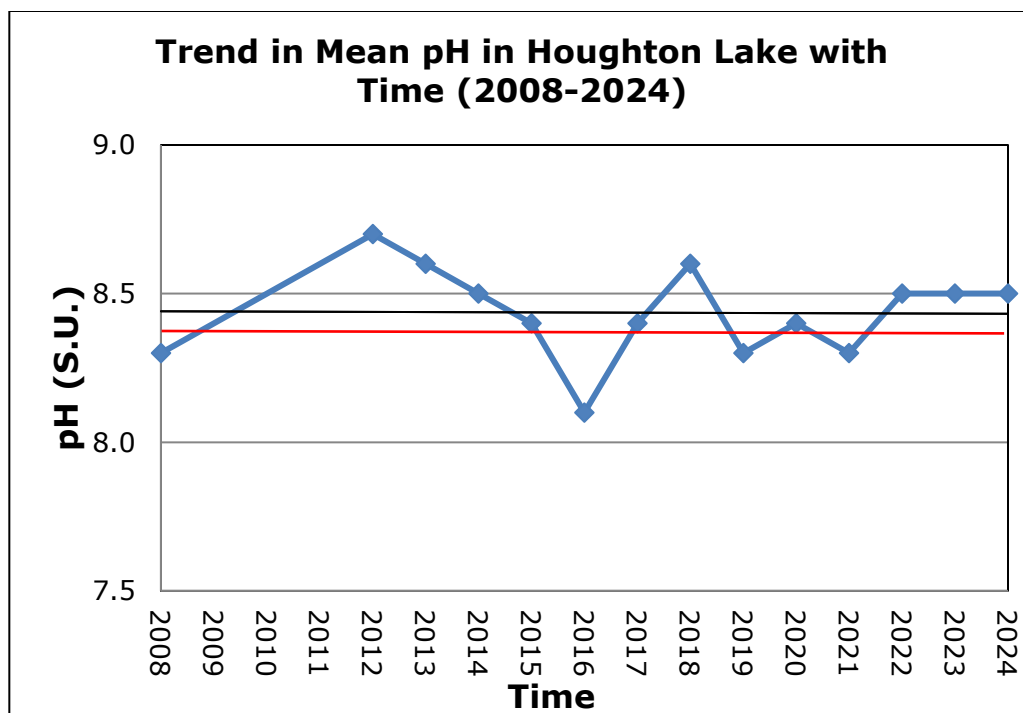


Figure 8. Changes in mean pH with time in Houghton Lake (2008-2024).

Conductivity

Conductivity is a measure of the number of mineral ions present in the water, especially those of salts and other dissolved inorganic substances and was measured with a calibrated Eureka Manta II® probe. Conductivity generally increases as the amount of dissolved minerals and salts in a lake increases and also increases as water temperature increases. The conductivity in Houghton Lake ranged from 235-287 $\mu\text{S}/\text{cm}$ on September 18, 2024. The conductivity of the canals ranged from 263-602 $\mu\text{S}/\text{cm}$ and the conductivity in the tributaries ranged from 262-280 $\mu\text{S}/\text{cm}$. Severe water quality impairments do not occur until values exceed 800 $\mu\text{S}/\text{cm}$ and are toxic to aquatic life around 1,000 $\mu\text{S}/\text{cm}$. The historic mean for the lake basin is 236 $\mu\text{S}/\text{cm}$ which is favorable, especially for a developed lake (Figure 9).

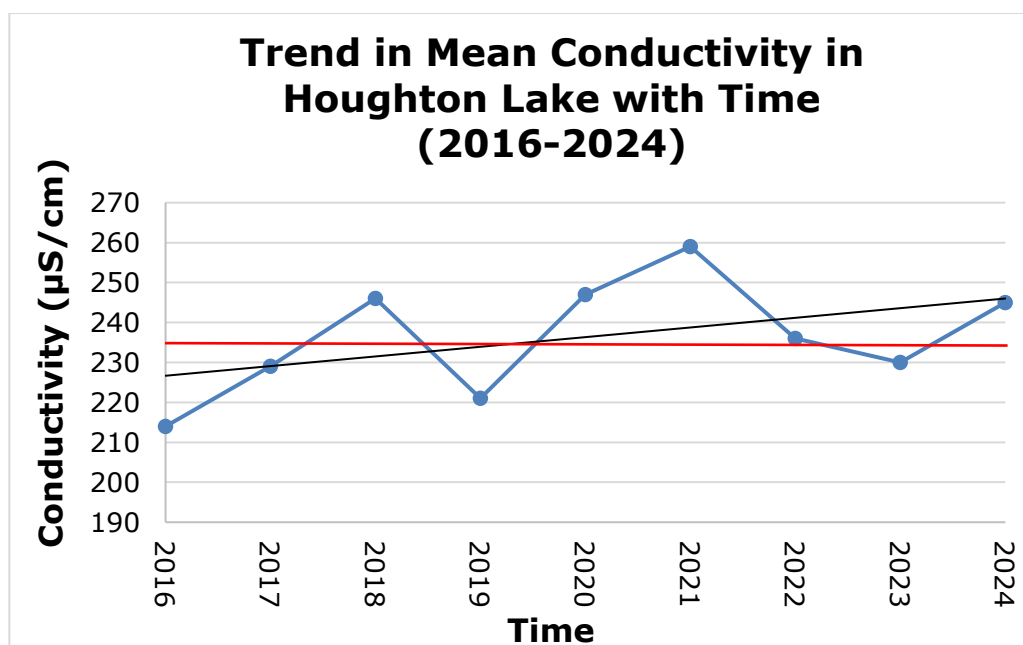


Figure 9. Changes in mean conductivity with time in Houghton Lake (2016-2024).

Chlorophyll-*a* and Algal Species Composition

Chlorophyll-*a* is a measure of the amount of green plant pigment present in the water, often in the form of planktonic algae. High chlorophyll-*a* concentrations are indicative of nutrient-enriched lakes. Chlorophyll-*a* concentrations greater than $6 \mu\text{g L}^{-1}$ are found in eutrophic or nutrient-enriched aquatic systems, whereas chlorophyll-*a* concentrations less than $2.2 \mu\text{g L}^{-1}$ are found in nutrient-poor or oligotrophic lakes. The mean chlorophyll-*a* concentration measured on September 18, 2024 (Figure 10) was $2.2 \mu\text{g L}^{-1}$ which was much lower than in recent years and may be attributed to less runoff and a dry climate. The historic mean chlorophyll-*a* concentration is $1.8 \mu\text{g L}^{-1}$ which is favorable and low.

The algal genera were determined from composite water samples collected over the deep basins of Houghton Lake in 2024 were analyzed with a compound Zeiss® bright field microscope. The genera present included the Chlorophyta (green algae): *Chlorella* sp., *Haematococcus* sp., *Ulothrix* sp., *Dictyosphaerium* sp., *Crucigenia* sp., and *Protococcus* sp.; *Spirogyra* sp., *Cladophora* sp., *Scenedesmus* sp., and *Pandorina* sp.; The Cyanophyta (blue-green algae): *Oscillatoria* sp., and *Microcystis* sp.; and the Bascillariophyta (diatoms): *Navicula* sp., *Cymbella* sp., *Synedra* sp., *Amphora* sp., *Fragillaria* sp., and *Tabellaria* sp.

The aforementioned species indicate a diverse algal flora and represent a good diversity of algae with an abundance of diatoms that are indicative of great water quality.

Blue-green algae have been historically problematic in the McKinley Canal system in recent years and Phoslock® treatments were conducted in 2021. In 2023, one of the Long Point canals (Figure 11) had a substantial blue-green algal bloom that was quickly treated with the algaecide SeClear® with good success. RLS will continue to evaluate the algal communities in the canals and make site-specific treatment recommendations when extreme blooms occur. The presence of blue-green algae in Houghton Lake requires careful and highly selective management of invasive aquatic plants since they compete with blue-green algae for nutrients in the water column.

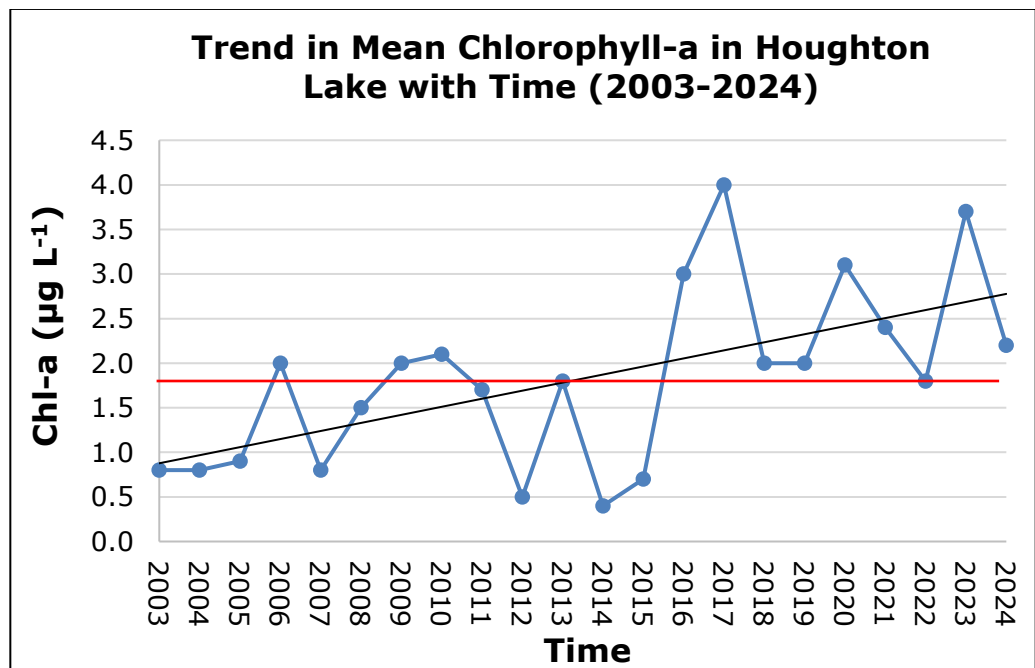


Figure 10. Changes in mean Chl-a with time in Houghton Lake (2003-2024).

The blue-green algae, *Microcystis* sp. has been the most prevalent algae in the canals, which is an indicator of poor water quality. The colonies are a few micrometers in diameter and are evenly distributed throughout a gelatinous matrix. Younger colonies are spherical and older ones are more irregularly shaped. There are numerous gas vesicles, and the algae can thrive at the surface with minimal photo-degradation (breaking down) by the sun. When the sunlight is excessive, the algae can break down and release toxins and lower the dissolved oxygen in the water column. The algae are the only type known to fix nitrogen gas into ammonia for growth. *Microcystis* has also been shown to overwinter in lake sediments (Fallon et al., 1981). In addition, it may thrive in a mucilage layer with sediment bacteria that can release phosphorus under anaerobic conditions (Brunberg, 1995). They assume a high volume in the water column (Reynolds, 1984) compared to diatoms and other single-celled green algae. The blue-green algae have been on the planet nearly 2.15 billion years and have assumed strong adaptation mechanisms for survival. In general, calm surface conditions will facilitate enhanced growth of this type of algae since downward transport is reduced. *Microcystis* may also be toxic to zooplankton such as *Daphnia* which was a zooplankton present in Houghton Lake and in most lakes (Nizan et al., 1986). Without adequate grazers to reduce algae, especially blue-greens, the blue-green population will continue to increase and create negative impacts to water bodies. Filamentous algae will also continue to increase in stagnant areas due to high nutrient levels in the lake.

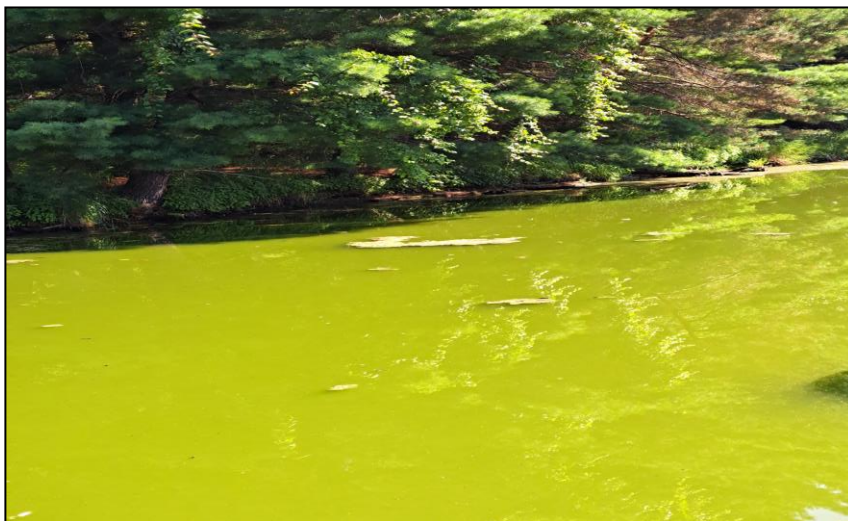


Figure 11. Localized blue-green algae bloom in a Long Point Canal (summer, 2023).

In 2023, a sample of algae on a rock was provided to RLS from a Houghton Lake Improvement Board member. The algae on the rock was identified with microscopy to be the green filamentous alga, *Cladophora* (Figure 12). *Cladophora* algae have become problematic in the Great Lakes and in many inland lakes and are considered a symptom of high nutrient loads processed by Zebra mussels (Higgins et al., 2008). *Cladophora* nearshore may become a nuisance because it produces strong sewage-like odors in nearshore areas, especially upon decay and may accumulate harmful bacteria such as *E. coli*.

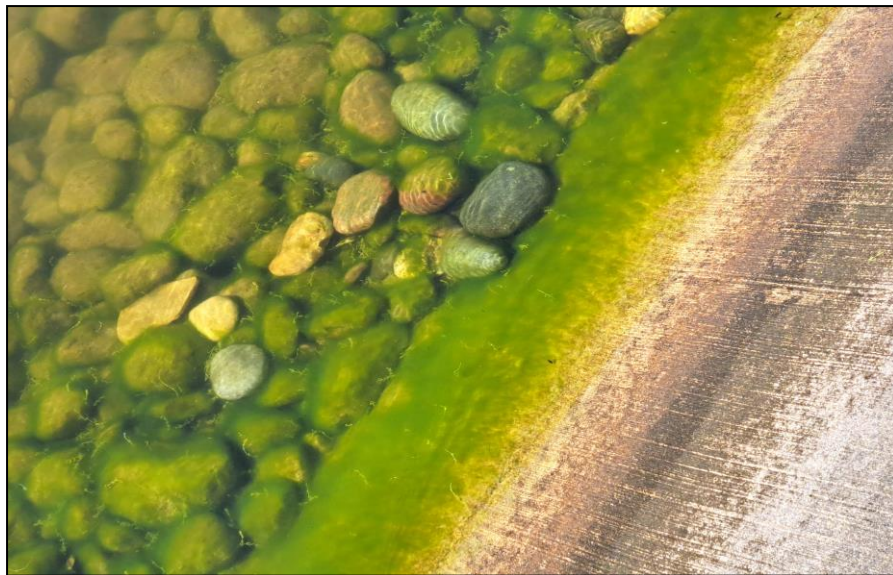


Figure 12. Green filamentous *Cladophora* algae growing on rocks and concrete in Houghton Lake (summer, 2023).

Given the ability of *Cladophora* to become a nuisance by fouling shoreline habitat and docks and hoists, RLS recommends a comprehensive baseline evaluation of *Cladophora* along the shoreline of Houghton Lake as well as additional measurements to possibly determine the likely causes of *Cladophora* in specific areas. RLS proposes to conduct this evaluation during the summer of 2025 and will issue a separate report of the findings and any needed mitigation recommendations.

Aquatic Vegetation Data (2024)

Status of Native Aquatic Vegetation in Houghton Lake

The native aquatic vegetation present in Houghton Lake is essential for the overall health of the lake and the support of the lake fishery. The June 4-21 2024 whole-lake survey using the GPS Point-Intercept method as in Figure 13 below determined that there were a total of 37 native aquatic plant species in Houghton Lake. These included 28 native submersed species, 2 floating-leaved species, and 7 emergent species. This indicates a very high biodiversity of aquatic vegetation in Houghton Lake that may change each year due to climate and germination conditions. The overall % cover of the lake by native aquatic plants has been historically low relative to the lake size due to the great mean depth and thus these plants should be protected. The overall aquatic vegetation biovolume has increased in recent years which is a positive sign that more low-growing native aquatic plants are thriving. The aquatic plant species found in the main open waters of the lake are shown below in Table 13. The aquatic vegetation biovolume for 2024 is displayed in Figure 14 below.

The EWM was significantly reduced in the Middle Grounds after the ProcettaCOR® treatment in 2019-20; however, the systemic herbicide triclopyr (Renovate 3® at doses of 4-5 gallons per acre) was used in 2021 since the EWM significantly rebounded in the Middle Grounds and use of the same product was not recommended in order to allow for product rotation. The Wild Rice population in the Middle Grounds is showing signs of re-establishment and thus treatments in this area will continue to include protective buffer zones to allow for this establishment and less herbicide will be needed with increased colonization by the Wild Rice.

The open waters of the lake are also quite diverse but have much less relative abundance than Middle Grounds. The most vegetated areas of open water in the lake include the southwest corner and Muddy Bay with some areas of density in North Bay. The milfoil in the North Bay has responded very well to treatments and native aquatic plants are increasing in that region as well.

Houghton Lake Survey Grid Points

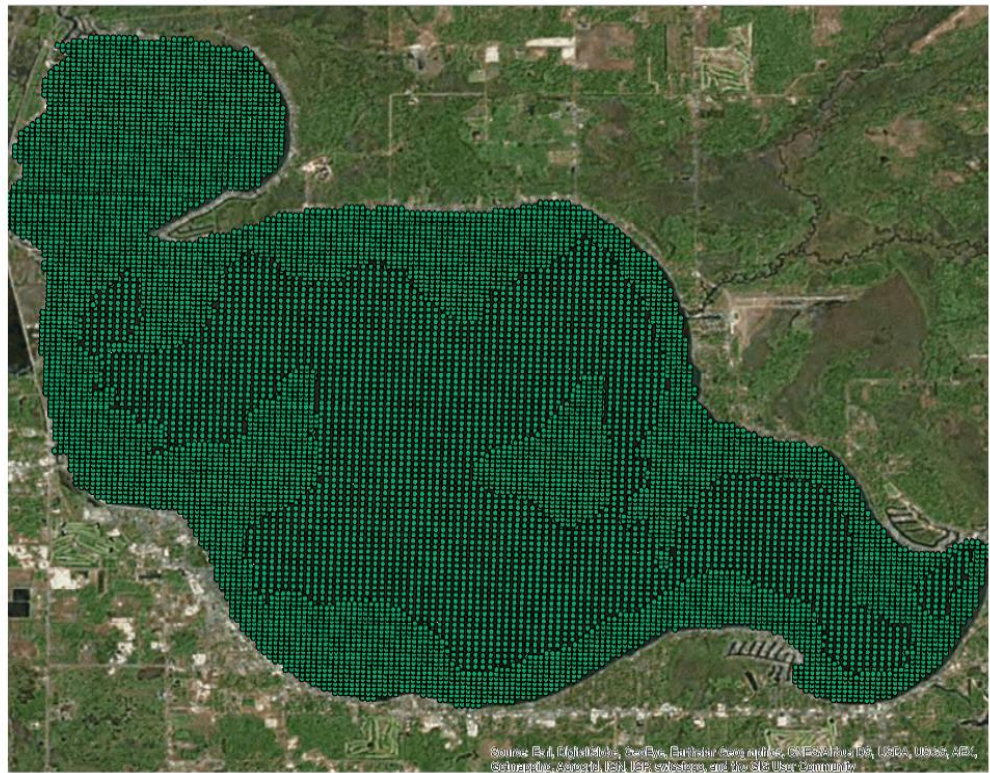


Figure 13. GPS Sampling Points in Houghton Lake (RLS).

Table 13. Native aquatic plant species relative abundance (frequency) in the main portion of the lake with vegetation present excluding canals (June 4-21, 2024).

Aquatic Plant Common Name	Aquatic Plant Latin Name	A level	B level	C level	D level	# Sites Found (% of total)
Muskgrass	<i>Chara vulgaris</i>	807	49	16	0	5.8
Sago Pondweed	<i>Stuckenia pectinata</i>	86	122	20	0	1.5
Flat-stem Pondweed	<i>Potamogeton zosteriformis</i>	208	59	11	1	1.9
Fern-leaf Pondweed	<i>Potamogeton robbinsii</i>	32	119	30	19	1.3
Thin-leaf Pondweed	<i>Potamogeton</i> spp.	2	1	1	0	0.03
American-leaf Pondweed	<i>Potamogeton nodosus</i>	12	33	0	0	0.3
Variable-leaf Pondweed	<i>Potamogeton gramineus</i>	782	258	193	168	9.3
White-stem Pondweed	<i>Potamogeton praelongus</i>	150	228	29	5	2.7
Clasping-leaf Pondweed	<i>Potamogeton richardsonii</i>	250	263	167	13	4.6
Illinois Pondweed	<i>Potamogeton illinoensis</i>	66	68	1	0	0.9
Large-leaf Pondweed	<i>Potamogeton amplifolius</i>	125	247	252	167	5.3
Floating-leaf Pondweed	<i>Potamogeton natans</i>	29	7	1	0	0.2
Small Pondweed	<i>Potamogeton pusillus</i>	94	233	88	14	2.9
Wild Celery	<i>Vallisneria americana</i>	7	6	3	0	0.1
Water Stargrass	<i>Zosterella dubia</i>	0	26	4	0	0.2
Northern Watermilfoil	<i>Myriophyllum sibiricum</i>	168	167	35	6	2.5
Variable-leaf Watermilfoil	<i>Myriophyllum heterophyllum</i>	1	4	3	1	0.06
Whorled Watermilfoil	<i>Myriophyllum verticillatum</i>	703	7	0	0	4.7
White Water-Crowfoot	<i>Ranunculus aquatilis</i>	44	0	0	0	0.3
Coontail	<i>Ceratophyllum demersum</i>	138	53	20	8	1.5
Aquatic Scorpion Moss	<i>Drepanocladus revolvens</i>	0	0	11	0	0.07
Common Waterweed	<i>Elodea canadensis</i>	210	38	71	1	2.1
Leafless Watermilfoil	<i>Myriophyllum tenellum</i>	19	16	0	0	0.2
Water Marigold	<i>Megalodonta beckii</i>	179	187	41	8	2.8
Bladderwort	<i>Utricularia vulgaris</i>	141	595	85	9	5.5
Southern Naiad	<i>Najas guadalupensis</i>	13	10	0	0	0.2
Brittle Naiad	<i>Najas minor</i>	38	29	17	4	0.6
Nitella	<i>Nitella</i> sp.	136	201	107	4	3.0
White Waterlily	<i>Nymphaea odorata</i>	63	116	82	2	1.7
Yellow Waterlily	<i>Nuphar variegata</i>	5	11	2	3	0.1
Arrowhead	<i>Sagittaria</i> sp.	188	18	1	3	1.4

Wild Iris	<i>Iris spp.</i>	0	0	32	0	0.2
Cattails	<i>Typha latifolia</i>	35	0	0	0	0.2
Swamp Loosestrife	<i>Decodon verticillatus</i>	5	0	1	0	0.04
Bulrushes	<i>Schoenoplectus acutus</i>	105	126	19	4	1.7
Pickerelweed	<i>Pontedaria cordata</i>	16	17	0	0	0.2
Wild Rice	<i>Zizania aquatica</i>	9	5	0	0	0.1

Note: There were a total of 15,038 points surveyed in the littoral zone of the main lake and 2,962 points in the canals. Approximately 67% of the main lake basin contained some aquatic vegetation growth.

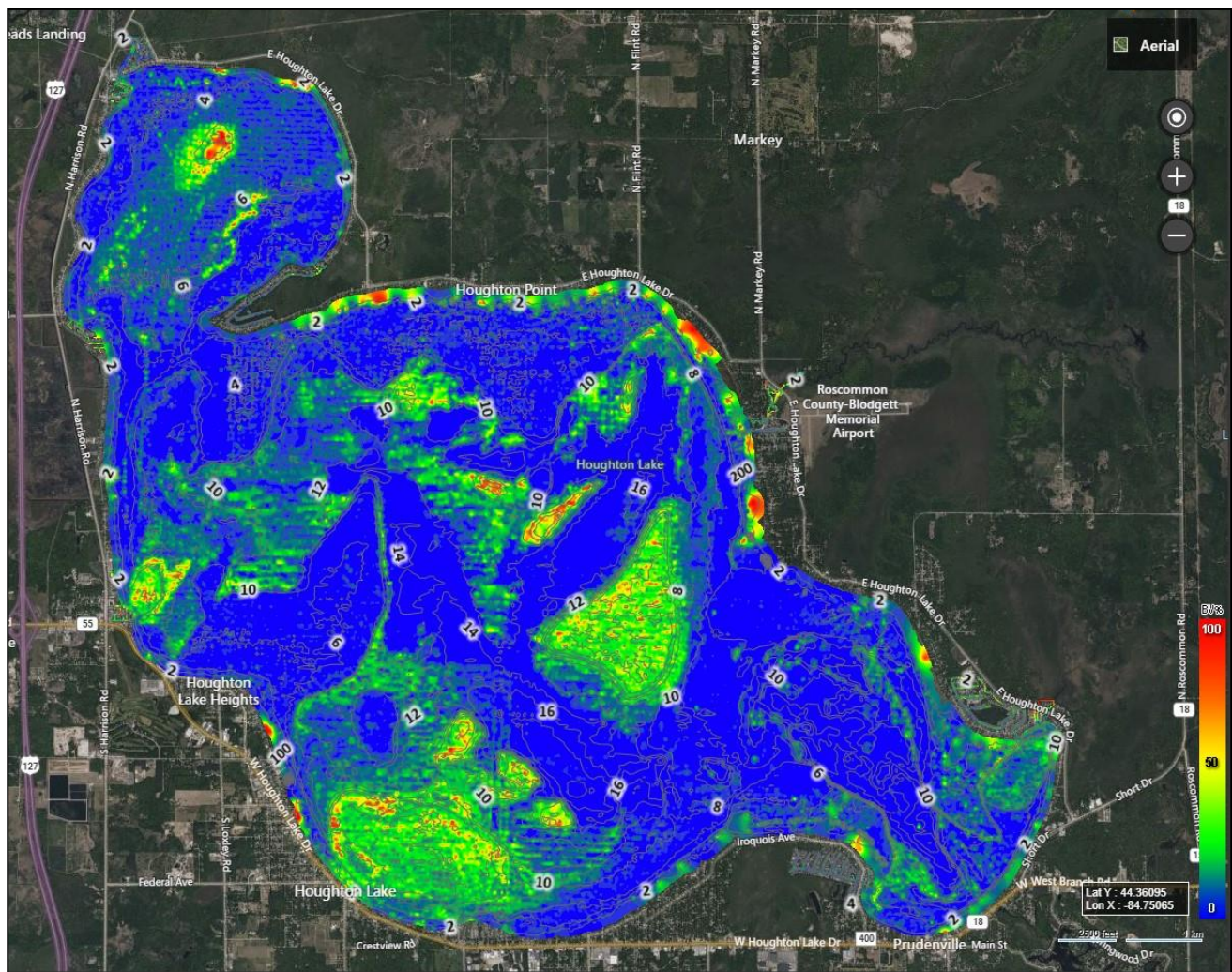


Figure 14. Aquatic vegetation biovolume scan and map of Houghton Lake in June, 2024 (RLS). NOTE: The blue color represents no vegetation present (previously this was displayed as blue and will be in the future); Red color represents tall, high-growing aquatic plants; Green color represents low-growing vegetation on the lake bottom such as *Chara*.

Status of Invasive (Exotic) Aquatic Plant Species

The amount of Eurasian Watermilfoil (Figure 15) present in Houghton Lake varies each year and is dependent upon climatic conditions, especially runoff-associated nutrients. The climate in 2024 consisted of low rainfall and abundant sunshine. Nutrient concentrations in the lake were lower than in recent years due to less runoff, so presumably less nutrients were available for submersed aquatic vegetation growth. However, an abundance of sunlight and increased Secchi transparency encouraged the growth of native aquatic plants throughout the lake basin. The majority of the treatment areas in 2023-2024 were in the canals as growth of invasives was limited in the main basin. The 2024 surveys revealed that a total of approximately 74.9 acres of milfoil were found throughout the main lake basin and necessitated treatment. These areas were treated beginning on May 21, 2024 and ending September 10, 2024. Treatments were conducted by PLM with oversight by RLS. Figures 15-18 display the invasives found in and around Houghton Lake in 2024. Figures 19-38 display critical treatment areas in 2024.

Table 14 displays the invasive species found in and around Houghton Lake in 2024 along with relative abundance and overall frequency.

Table 15 below shows the 2024 treatment history with the amounts of contact and systemic herbicides used in Houghton Lake for milfoil/nuisance growth treatments and in some canals the use of contacts for extremely dense vegetation.

Table 14. Invasive aquatic plant species relative abundance (frequency) in the main portion of the lake with vegetation present excluding canals (June 4-21, 2024).

Aquatic Plant Common Name	Aquatic Plant Latin Name	A level	B level	C level	D level	# Sites Found (% of total)
Starry Stonewort	<i>Nitellopsis obtusa</i>	33	94	20	2	1.0
Eurasian Watermilfoil (Hybridized)	<i>Myriophyllum spicatum</i> var. <i>sibiricum</i>	388	80	23	0	3.3
Curly-leaf Pondweed	<i>Potamogeton crispus</i>	315	42	14	0	2.5
Purple Loosestrife	<i>Lythrum salicaria</i>	48	133	0	0	1.2

Table 15. Aquatic herbicide treatments conducted in Houghton Lake canals and the lake basin in 2024.

Herbicide Treatment Date in Lake/Canals	Aquatic Plants Targeted	Acres Treated	Aquatic Herbicides Used	Total Cost
May 21, 2024	EWM/CLP	48.4	ProcellaCOR® (6 PDU); Diquat (1gal/ac); Flumioxazin (200 ppb)	\$24,842.78
June 20, 2024	EWM	188.2	Renovate 3® (8 gal/ac); Diquat (0.5 gal/ac); ProcellaCOR® (6-8 PDU); diquat (1 gal/ac); Flumioxazin	\$100,626.52
July 23, 2024	EWM/Starry	12.9	Diquat (1 gal/ac); ProcellaCOR® (6 PDU); SeClear G®	\$7,262.20
September 10, 2024	EWM/Starry	82.6	SeClear G; Diquat (100 ppb)	\$21,059.06
TOTALS	—	332.1	--	\$153,790.56



Figure 15. Eurasian Watermilfoil with seed head and lateral branches.



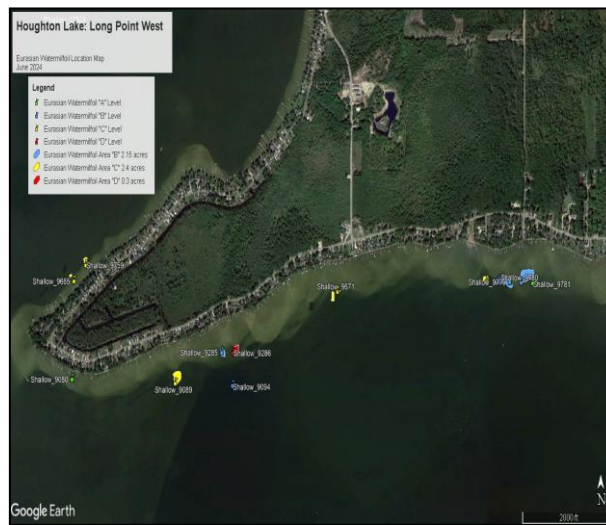
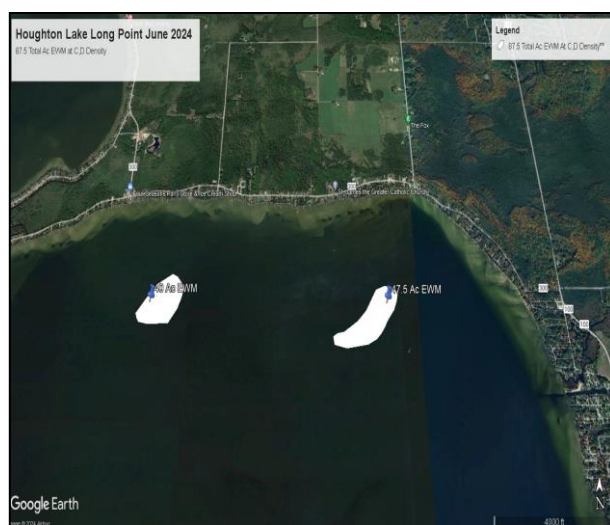
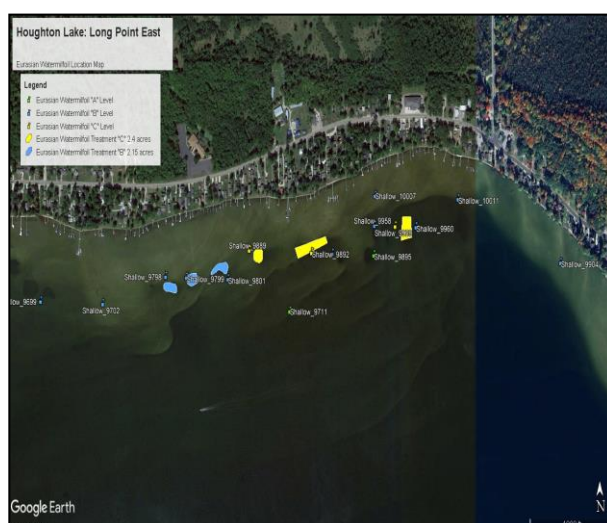
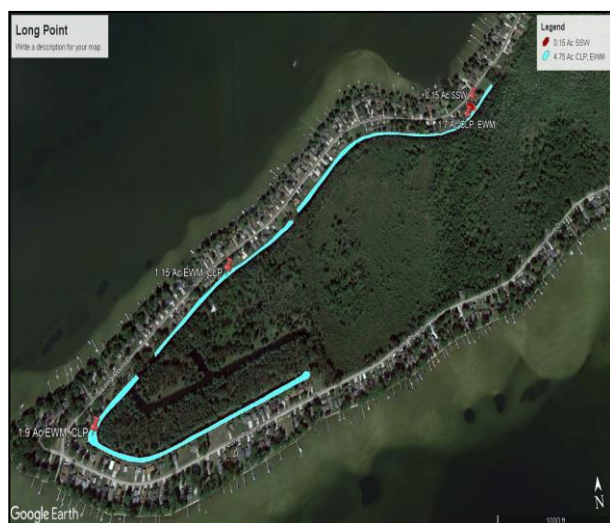
Figure 16. Curly-leaf Pondweed with wavy leaf margins.



Figure 17. Starry Stonewort with attached star-shaped bulbils.



Figure 18. Emergent Purple Loosestrife.



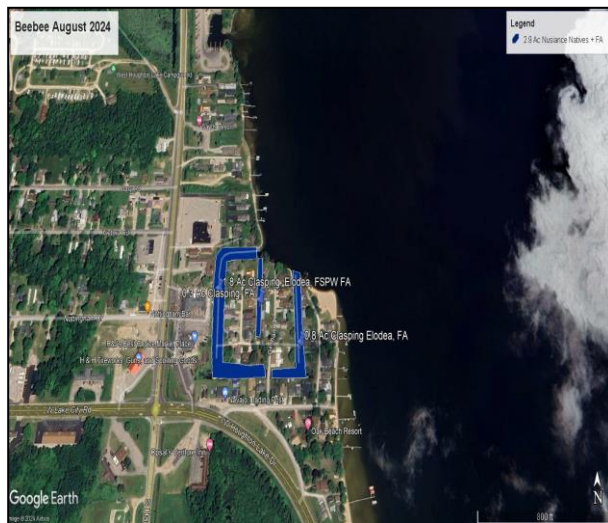


Figure 23. Treatment Area (Beebe Canals)



Figure 24. EWM/SS/CLP Treatment Area (Birch Crest Muskegon River)



Figure 25. CLP Treatment (Chippewa)

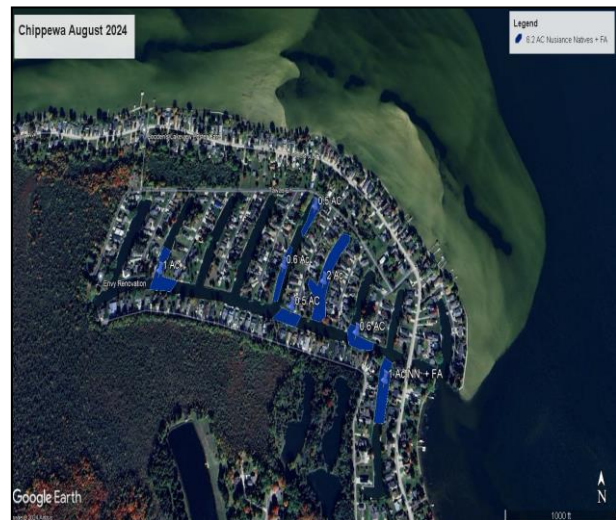
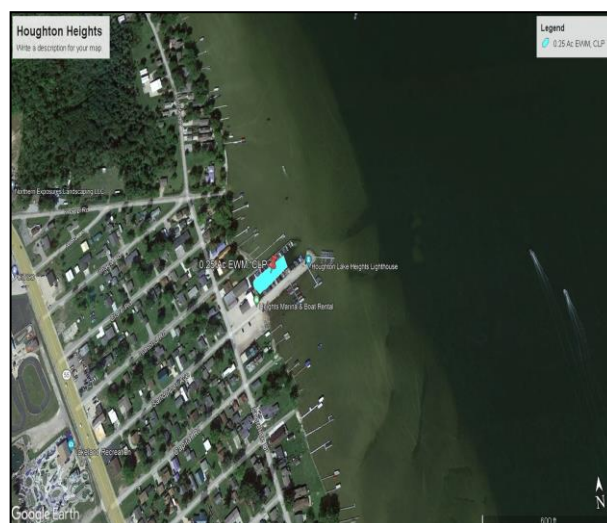
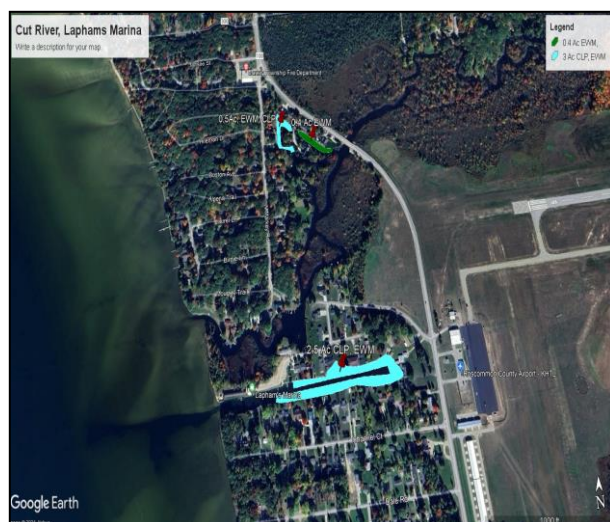
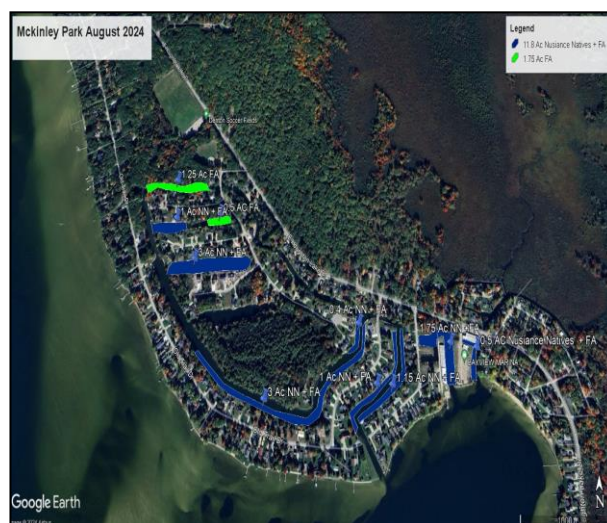
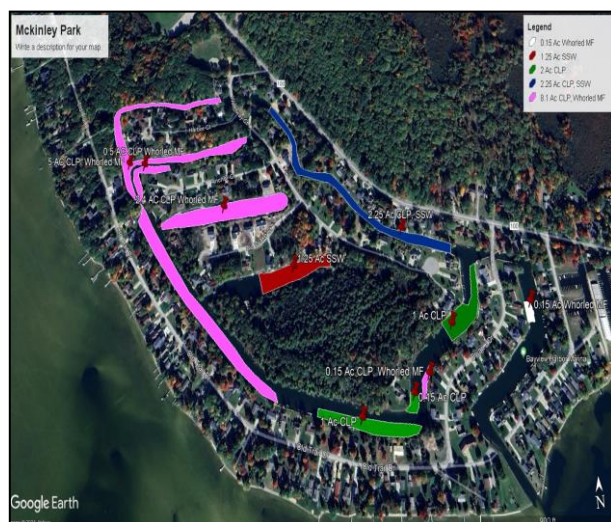


Figure 26. Nuisance Natives Treatment Area (Chippewa)



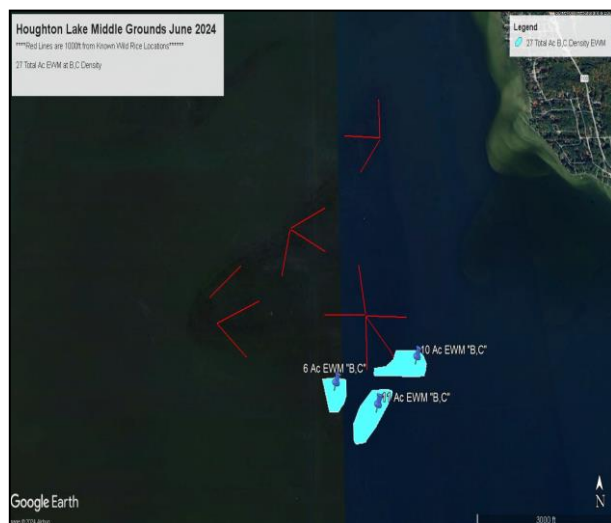


Figure 31. EWM Treatment Areas (Middle Grounds)

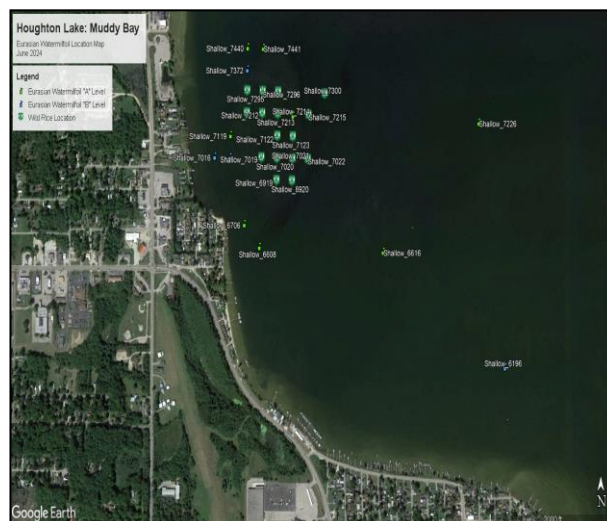


Figure 32. EWM Treatment areas (Muddy Bay, excl. Wild Rice Areas)

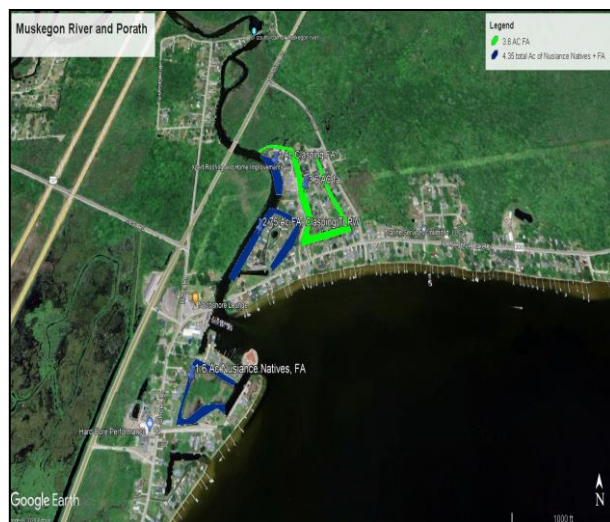


Figure 33. Nuisance Natives and dense Algae Treatment (Muskegon River)

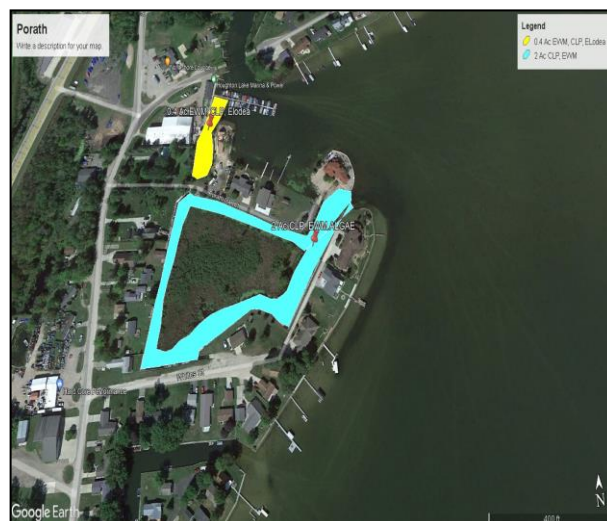


Figure 34. EWM/CLP Treatment Area (Porath Canal)

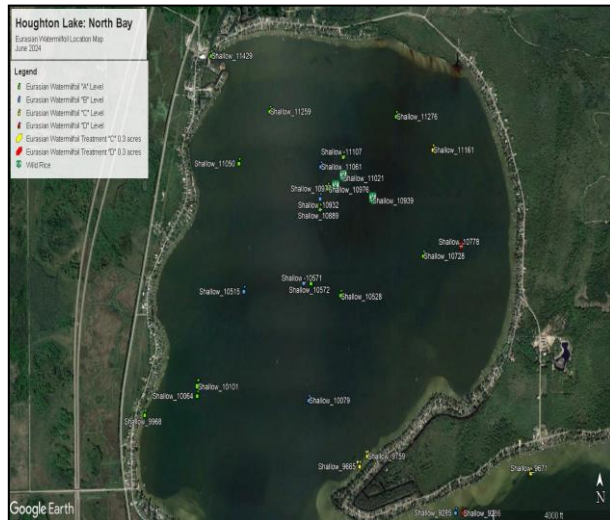


Figure 35. EWM Treatment Areas (North Bay)



Figure 36. EWM Treatment areas (Southwest Bay); CLP not treated

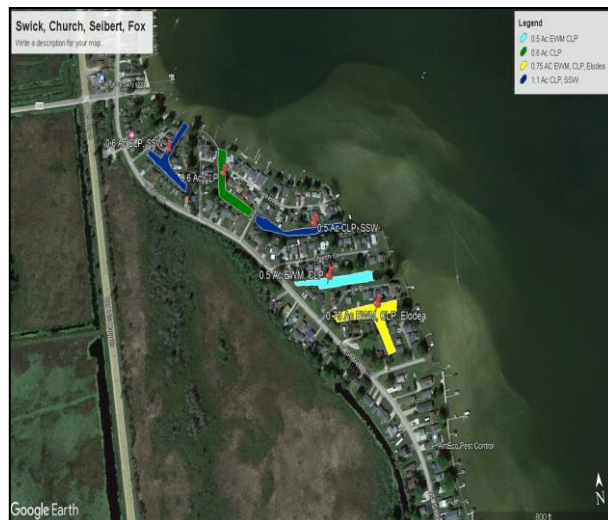


Figure 37. EWM/CLP/SSW Treatment (Swift, Church, Siebert, Fox Canals)

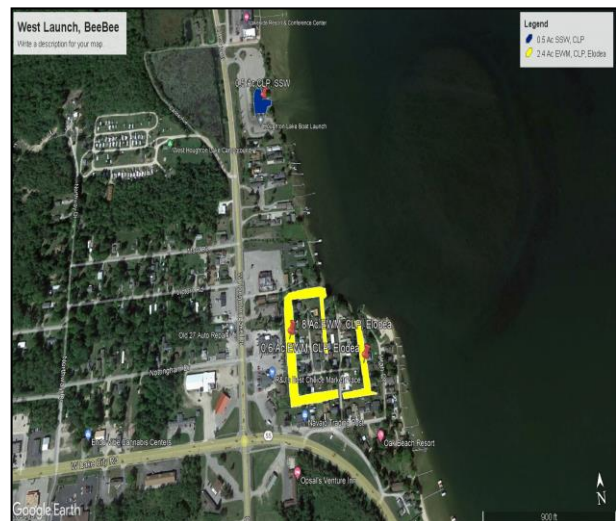


Figure 38. EWM/CLP Treatment Area (West Launch Beebe Canal)

Houghton Lake Sediment Aquatic Macroinvertebrates

RLS scientists collected sediment macroinvertebrate communities from the North Bay, Central Basin, and South Bay on October 4, 2024 so they may be compared to earlier sample data and also determine the existing biodiversity of taxa that contribute to the ecological balance of Houghton Lake. Tables 16-19 list all of the aquatic macroinvertebrates found during the 2024 sampling.

A previous study on the Houghton Lake macroinvertebrate community determined that the total number of macroinvertebrate taxa declined from 19 in 1973 to 9 by 1995-1996. The October 4, 2024 samples demonstrated 11 different taxa in the lake sediments and this number is likely to fluctuate among seasons due to changes in environmental and climatic conditions. Thus, future preservation is important since these organisms support the lake food chain and fishery. In 2024, the North Basin had the highest counts. Taxa found in the samples included:

1. Pond snails
2. Mayfly larvae
3. Square gill may fly larvae
4. Wheel snails
5. Dragonfly larvae
6. Midge larvae
7. Isopods
8. Flatworms
9. Crane fly larvae
10. Damselfly larvae
11. Long horned caddisflies

Table 16. Houghton Lake sediment macroinvertebrate sampling data from the North Bay (October 4, 2024).

Sample 1	Grab	Order	Family/Genus	Number	Common name
		Caecidotea	Asellidae	5	Isopods
		Ephemeroptera	Ephemeridae	8	Mayfly larvae
		Planaria	Planariidae	1	Flatworms
		Diptera	Chironomidae	28	Midge larvae
		Gastropoda	Physidae	3	Pond snails
		Odonata	Lestidae	2	Damselfly
		Gastropoda	Planorbidae	16	Wheel snails
			Total	63	
Sample 2	Grab	Order	Family/Genus	Number	Common Name
		Gastropoda	Physidae	13	Pond snails
		Ephemeroptera	Ephemeridae	1	Mayfly larvae
		Caecidotea	Asellidae	5	Isopods
		Diptera	Tipulidae	2	Crane fly larvae
		Gastropoda	Planorbidae	10	Wheel snails
		Odonata	Aeshniidae	1	Dragonfly larvae
		Diptera	Chironomidae	19	Midge larvae
			Total	51	

Table 17. Houghton Lake sediment macroinvertebrate sampling data from the Central Basin (October 4, 2024).

Sample 1	Grab	Order	Family/Genus	Count	Common name
		Diptera	Tipulidae	2	Crane fly larvae
		Ephemeroptera	Ephemeridae	3	Mayfly larvae
		Diptera	Chironomidae	17	Midge larvae
		Caecidotea	Asellidae	11	Isopods
		Gastropoda	Planorbidae	13	Wheel snails
			Total	46	
Sample 2	Grab				
		Gastropoda	Physidae	16	Pond snails
		Ephemeroptera	Ephemeridae	2	Mayfly larvae
		Gastropoda	Coenagrionidae	2	Damselfly larvae
		Gastropoda	Planorbidae	24	Wheel snails
		Diptera	Chironomidae	8	Midge larvae
		Diptera	Tipulidae	2	Crane fly larvae
			Total	54	

Table 18. Houghton Lake sediment macroinvertebrate sampling data from the South Basin (October 4, 2024).

Sample 1	Grab	Order	Family/Genus	Number	Common name
		Tricoptera	Leptoceridae	4	Long Horned caddisflies
		Gastropoda	Planorbidae	10	Wheel Snails
		Ephemeroptera	Ephemeridae	4	Mayfly larvae
		Diptera	Chironomidae	19	Midge larvae
		Gastropoda	Physidae	2	Pond snails
		Ephemeroptera	Caenidae	2	Square gill Mayfly
			Total	41	
Sample 2	Grab				
		Gastropoda	Physidae	11	Pond snails
		Ephemeroptera	Ephemeridae	2	Mayfly larvae
		Gastropoda	Planorbidae	8	Wheel snails
		Diptera	Chironomidae	17	Midge larvae
		Odonata	Lestidae	4	Damselfly larvae
			Total	42	

2024 Wild Rice Restoration Update and 2025 Restoration Recommendations:

Previously, RLS accompanied Dr. Scott Herron from Ferris State University on September 22, 2020 to a 50-acre area of Muddy Bay to complete the initial planting of Wild Rice in that region. Conditions in the Muddy Bay region were ideal for Wild Rice with shallow depths and a highly organic bottom substrate. A total of 108 geo-referenced GPS points were recorded and randomly selected from within the 50-acre area for data recording. A total of 22 bags of Wild Rice were carefully hand-tossed into the water and the seeds made fast contact with the lake bottom. A follow-up survey of this seeded area occurred on May 17, 2021 to carefully monitor the efficacy of the Wild Rice planting. Table 18 below displays the data collected which includes the presence of 7 native submersed aquatic plants in addition to the emergent Wild Rice. Wild Rice was the most dominant aquatic plant present, occupying around 98.8% of the sampling sites. This was a very successful germination. A second survey (Table 20) was conducted on May 13, 2022 and determined that the Wild Rice population had declined to 12.5% of the sampling sites and invasive hybrid Eurasian Watermilfoil occupied 37.5% of the sampling sites. The May 26, 2023 (Table 21) revealed 8.8% of the sampling sites contained Wild Rice and milfoil had declined to 13.8%. In May 2024 (Table 22), the Wild Rice in Muddy Bay had naturally increased to 11.3% and the milfoil was around 18.8%.

During the summer of 2023, Wild Rice began to naturally re-colonize the Middle Grounds with over 4 acres of active growth (Figure 39). Many of the plants developed healthy seeds (Figure 40) where some dropped to the lake bottom and others were consumed by waterfowl. The North Bay is beginning to demonstrate some natural growth (Figure 41). That area is a favorable candidate for seeding in the future if sources are available.

In 2024, the Middle Grounds had a population of around 40 acres of natural Wild Rice growth (Figure 42). This was a substantial increase from 2023 and is likely due to strong reductions in EWM and lower water levels.

Figures 43-44 show the observed 2024 growth of Wild Rice in the Cut River region.

Muddy Bay is showing some sustained growth since its original seeding in 2022 but may be prone to excessive scouring from ice and also intense winds due to its position in the lake basin relative to the fetch.

In 2023, in addition to the surveys conducted, RLS also collected a total of 12 sediment samples using an Ekman hand dredge in the lake basin and in areas with Wild Rice present. These sediment samples were analyzed for sediment percentage of organic matter (carbon) and nutrients such as phosphorus and inorganic nitrogen. The results indicated that there were no significant differences in mean sediment organic matter between the Wild Rice and main basin sites (Figures 45-49). However, the Wild Rice sites contained significantly higher mean concentrations of ammonia and phosphorus in the sediments. This indicates rationale for the presence of Wild Rice beds in specific areas of Houghton Lake and is useful for consideration of possible future planting sites for successful germination and survival. This research was presented at the October, 2023 NALMS conference in Erie, PA. It is currently in the process of being prepared and submitted for publication in a peer-reviewed scientific journal and was recently presented to the Michigan Wild Rice Initiative Monitoring and Restoration Sub-Committee during November, 2024.



Figure 39. Rigorous natural Wild Rice growth emergent in the Middle Grounds of Houghton Lake (summer, 2023).



Figure 40. A mature and seeding Wild Rice plant in the Middle Grounds of Houghton Lake (summer, 2023).

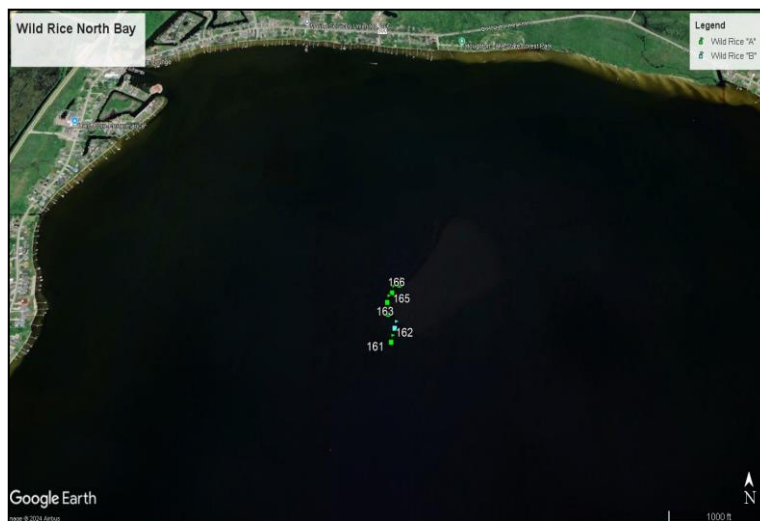


Figure 41. Natural Wild Rice growth emergent in the North Bay of Houghton Lake (summer, 2024).

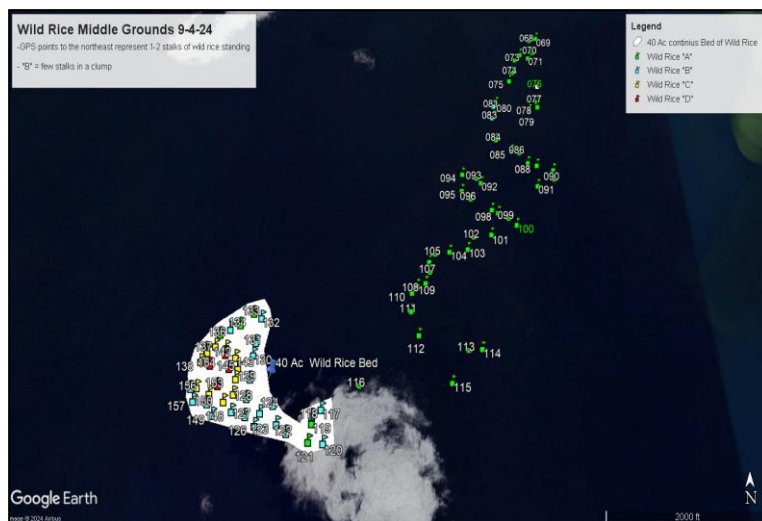


Figure 42. Natural Wild Rice growth emergent in the Middle Grounds of Houghton Lake (summer, 2024).



Figure 43. Natural Wild Rice growth emergent in the Cut River Region of Houghton Lake (summer, 2024).

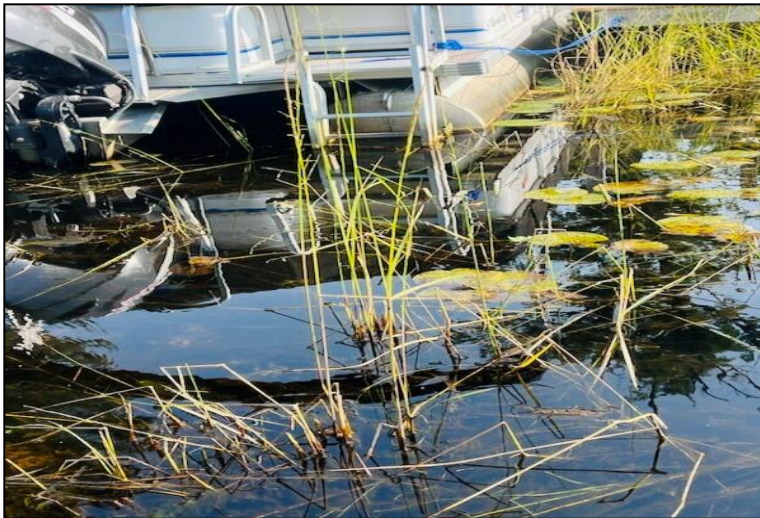


Figure 44. Natural Wild Rice growth emergent in the Cut River Region of Houghton Lake (summer, 2024).

Table 19. Aquatic vegetation survey data for the Wild Rice seeded area of Muddy Bay on May 17, 2021.

Aquatic Plant Common Name	Aquatic Plant Latin Name	A level	B level	C level	D level	# Sites Found (% of N=80 sites)
Muskgrass	<i>Chara vulgaris</i>	2	2	0	0	5.0
Curly-leaf Pondweed	<i>Potamogeton crispus</i>	21	19	9	0	61.3
Flat-stem Pondweed	<i>Potamogeton zosteriformis</i>	1	0	0	0	1.3
White-stem Pondweed	<i>Potamogeton praelongus</i>	1	0	0	0	1.3
Illinois Pondweed	<i>Potamogeton illinoensis</i>	12	17	0	0	36.3
Common Waterweed	<i>Elodea canadensis</i>	3	1	0	0	5.0
Southern Naiad	<i>Najas guadalupensis</i>	0	1	0	0	1.3
Wild Rice	<i>Zizania palustris</i>	4	48	27	0	98.8

Table 20. Aquatic vegetation survey data for the Wild Rice seeded area of Muddy Bay on May 13, 2022.

Aquatic Plant Common Name	Aquatic Plant Latin Name	A level	B level	C level	D level	# Sites Found (% of N=80 sites)
Muskgrass	<i>Chara vulgaris</i>	33	11	0	0	55.0
Hybrid Watermilfoil	<i>Myriophyllum spicatum</i> var. <i>sibiricum</i>	15	15	0	0	37.5
Fern-leaf Pondweed	<i>Potamogeton robbinsii</i>	1	0	0	0	1.3
Common Waterweed	<i>Elodea canadensis</i>	1	0	0	0	1.3
Southern Naiad	<i>Najas guadalupensis</i>	0	1	0	0	1.3
Wild Rice	<i>Zizania palustris</i>	10	0	0	0	12.5

Table 21. Aquatic vegetation survey data for the Wild Rice seeded area of Muddy Bay on May 26, 2023.

Aquatic Plant Common Name	Aquatic Plant Latin Name	A level	B level	C level	D level	# Sites Found (% of N=80 sites)
Muskgrass	<i>Chara vulgaris</i>	12	6	1	0	23.8
Curly-leaf Pondweed	<i>Potamogeton crispus</i>	6	2	2	1	13.8
Illinois Pondweed	<i>Potamogeton illinoensis</i>	18	1	4	0	28.8
Hybrid Watermilfoil	<i>Myriophyllum spicatum</i> var. <i>sibiricum</i>	8	2	1	0	13.8
Fern-leaf Pondweed	<i>Potamogeton robbinsii</i>	1	1	0	0	2.5
Common Waterweed	<i>Elodea canadensis</i>	2	1	0	0	3.8
Southern Naiad	<i>Najas guadalupensis</i>	6	1	2	1	12.5
Wild Rice	<i>Zizania palustris</i>	3	3	1	0	8.8

Table 22. Aquatic vegetation survey data for the Wild Rice seeded area of Muddy Bay on May 22, 2024.

Aquatic Plant Common Name	Aquatic Plant Latin Name	A level	B level	C level	D level	# Sites Found (% of N=80 sites)
Muskgrass	<i>Chara vulgaris</i>	18	3	2	0	28.8
Curly-leaf Pondweed	<i>Potamogeton crispus</i>	8	7	5	0	25.0
Illinois Pondweed	<i>Potamogeton illinoensis</i>	10	3	0	0	16.3
Hybrid Watermilfoil	<i>Myriophyllum spicatum var. sibiricum</i>	9	4	0	1	17.8
Fern-leaf Pondweed	<i>Potamogeton robbinsii</i>	3	2	0	0	6.3
Common Waterweed	<i>Elodea canadensis</i>	8	3	1	0	15.0
Southern Naiad	<i>Najas guadalupensis</i>	9	5	1	0	18.8
Wild Rice	<i>Zizania palustris</i>	5	2	2	0	11.3

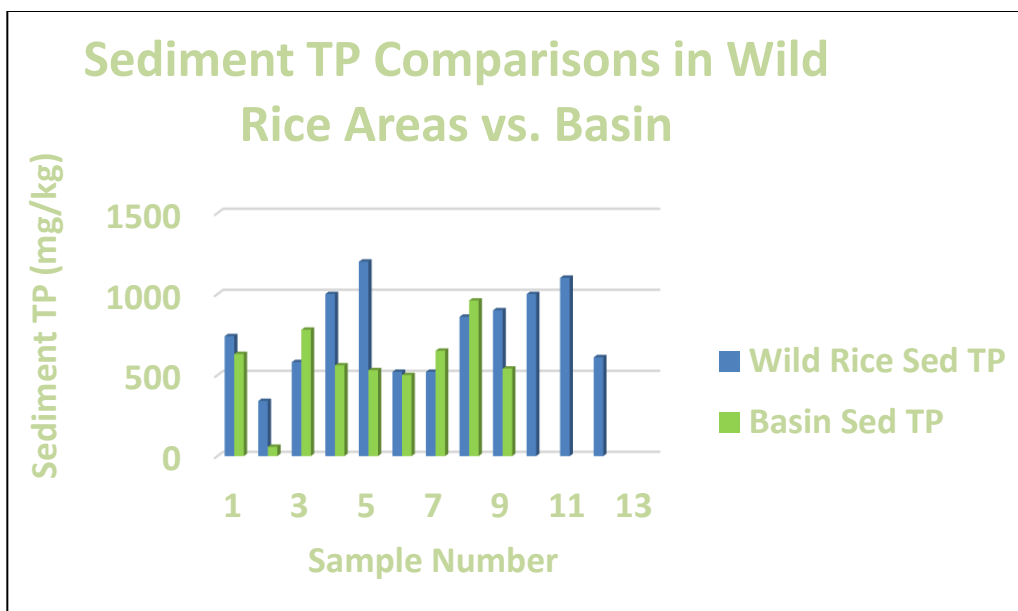


Figure 45. Differences in sediment TP in Wild Rice areas relative to lake basin sites (2023).

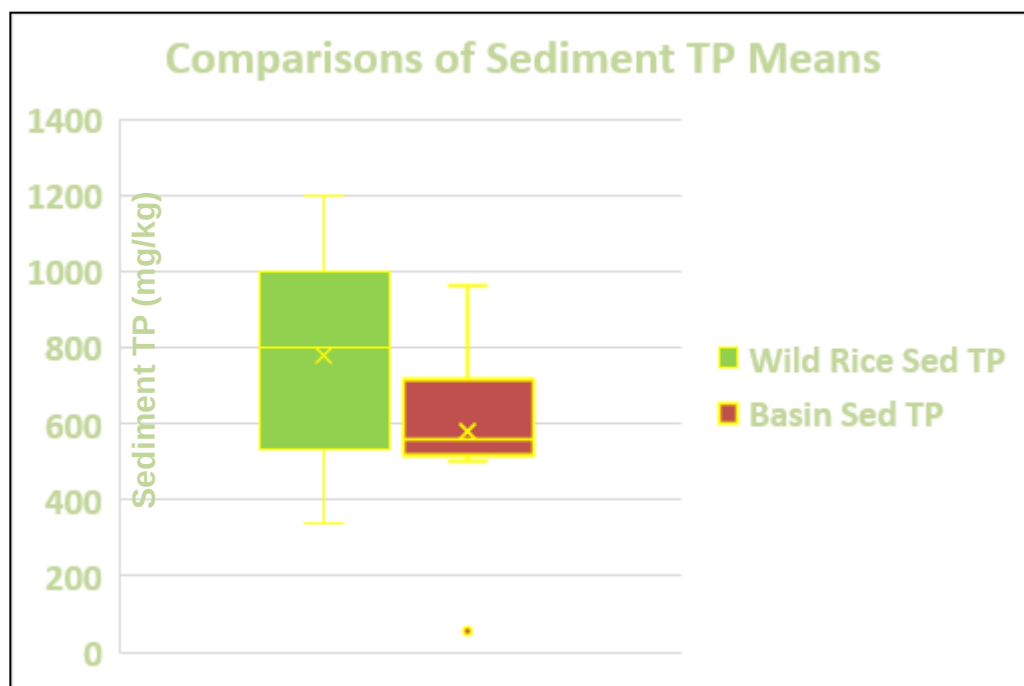


Figure 46. Differences in sediment TP means in Wild Rice areas relative to lake basin sites (2023).

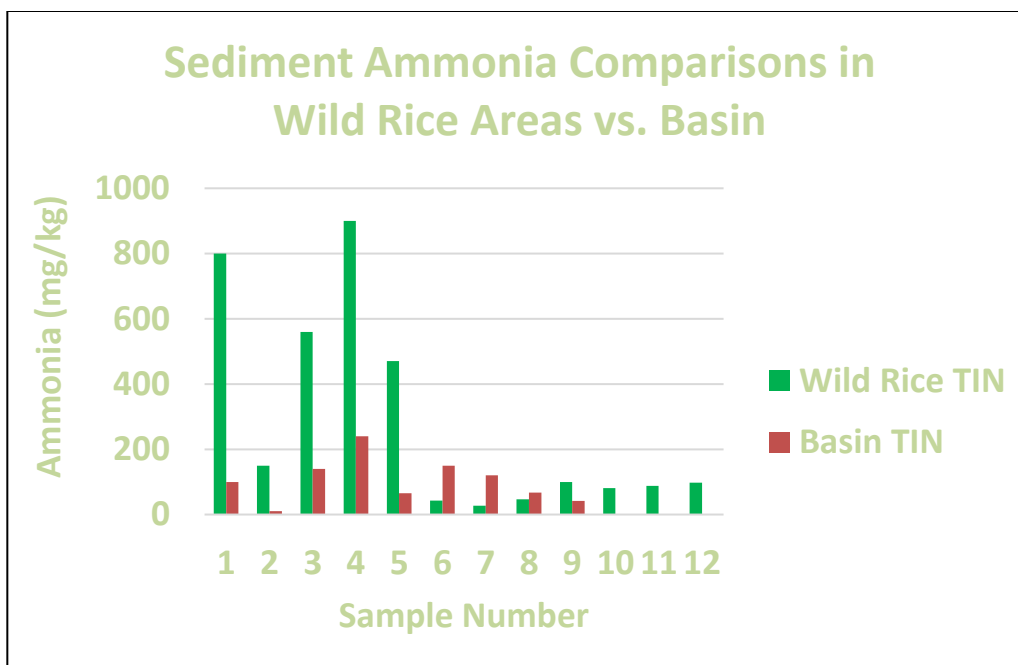


Figure 47. Differences in sediment ammonia in Wild Rice areas relative to lake basin sites (2023).

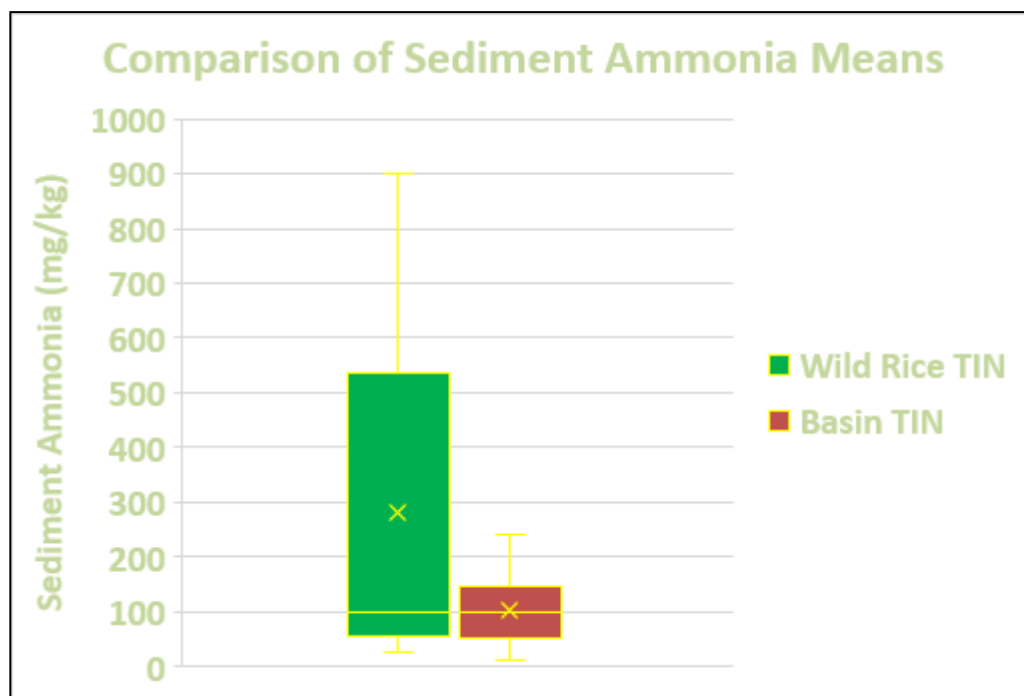


Figure 48. Differences in sediment ammonia means in Wild Rice areas relative to lake basin sites (2023).

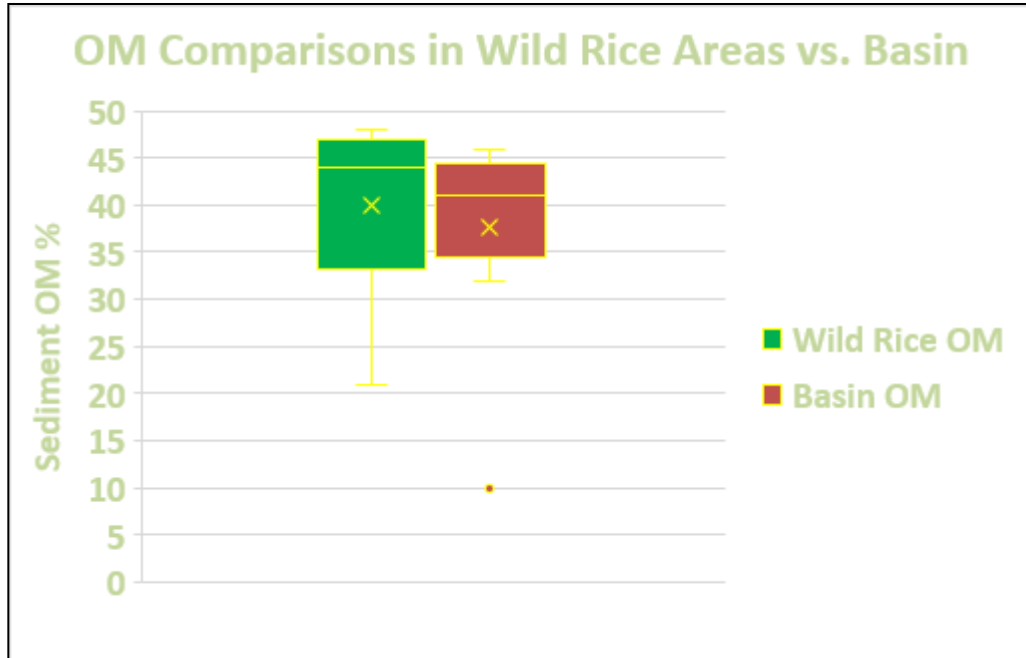


Figure 49. Differences in sediment organic matter (carbon) means in Wild Rice areas relative to lake basin sites (2023).

Management Recommendations for 2024-2026

As previously recommended in the 2022 Houghton Lake Improvement Feasibility Study report, RLS has recommended the following management activities for 2022-2026 as critical components for a continuing lake improvement (management) program. The primary and secondary goals of these management activities are shown below in Table 23. Ongoing proposed costs for 2024-2026 are displayed below in Table 24.

1. Whole-lake Aquatic Vegetation Surveys & Scans:

Continued aquatic vegetation surveys are needed to determine the precise locations of Eurasian Watermilfoil (EWM) Curly-leaf Pondweed (CLP), Starry Stonewort, or other problematic invasives in or around Houghton Lake and in the canals as in past years. These surveys will include a whole lake inventory in late June-early July 2022-2026 and partial surveys post-treatment as needed. The canals and Middle Grounds as well as the southwest regions of the lake may require earlier surveys beginning in mid to late May, depending on climatic conditions. Scientists from RLS will be present to oversee all aquatic herbicide treatments in 2025 as in previous years. Treatment results will then be compared with previous years in the 2025 annual lake report.

2. Aquatic Herbicide Treatments for Invasive Species in the Main Lake and Canals:

Due to the relative scarcity of native aquatic vegetation in the main basin of Houghton Lake, the treatment of these species with aquatic herbicides is not recommended and re-colonization of the lake by these species is a major goal for the current Houghton Lake management plan. The exception would be in the canals where often a mixture of invasives, nuisance natives, and dense algae can be found. The plan for 2022-2026 includes the use of high doses of systemic aquatic herbicides (such as triclopyr, 2,4-D, and/or ProcellaCOR® for the milfoil that may be present. Doses will be dependent upon the permit requirements as well as the size and density of the weed beds. Lower doses are used in the sensitive Middle Grounds area and in any areas where RLS finds Wild Rice during the whole-lake survey.

Additionally, RLS will continue to individually evaluate previously treated ProcellaCOR® treatment areas and any new areas that may be added with that product.

Thus far, the ProcellaCOR® product has proven to be a very effective herbicide for controlling the density and relative abundance of EWM without reducing favorable native aquatic plant species. The use of it may be alternated with other products to allow for reduced probability of tolerance. It has been particularly favorable as a product in rotation with triclopyr in the Middle Grounds where Wild Rice is showing strong natural germination and growth. RLS will also discuss the possibility of a test area with the newer product Galleon®, which can systemically treat Curly-leaf Pondweed in areas where it may grow together with EWM for maximum control of both plants.

3. Phoslock® Treatment of Select Canals:

The presence of toxic blue-green algal blooms is a threat to the health of some canals and pets that may drink from them. RLS recommended and evaluated the innovative product Phoslock® on the McKinley Canal System in 2021. Overall, the product showed significant reductions in blue-green algal concentrations. In 2023, the Long Point canal had a significant blue-green algae bloom as was promptly treated with SeClear® with positive visible and measurable results. RLS will continue to evaluate all canals that may need these treatments in future years.

4. Benthic Barriers and Weed Rollers:

Both of these technologies are simple to install and may be used in nearshore areas to reduce and/or prevent germination of submersed aquatic vegetation in beach areas and around docks. They act to reduce germination of all aquatic plants and lead to a local area free of most aquatic vegetation. Benthic barriers may come in various sizes between 100-400 feet in length. They are anchored to the lake bottom to avoid becoming a navigation hazard. The implementation of a benthic barrier mat requires a minor permit from EGLE which can cost around \$50-\$100. The cost of the barriers varies among vendors but can range from \$100-\$1,000 per mat. Benthic barrier mats can be purchased online at: www.lakemat.com or www.lakebottomblanket.com. The efficacy of benthic barrier mats has been studied by Laitala et al. (2012) who report a minimum of 75% reduction in invasive milfoil in the treatment areas. Lastly, benthic barrier mats should not be placed in areas where fishery spawning habitat is present and/or spawning activity is occurring.

Weed Rollers are electrical devices which utilize a rolling arm that rolls along the lake bottom in small areas (usually not more than 50 feet) and pulverizes the lake bottom to reduce germination of any aquatic vegetation in that area. They can be purchased online at: www.crary.com/marine or at: www.lakegroomer.net.

5. Mechanical Harvesting in Select Areas:

The use of a mechanical harvesting machine may continue to be needed for problem areas with extremely dense aquatic vegetation such as the Beebe Canal or other canals. This method is often preferred when the quantity of biomass is so large that contact herbicides may cause an unacceptable decline in dissolved oxygen in the water column upon rapid decay. This may not be needed every year but will be evaluated on an as-needed basis. Permission is obtained from John Hanes with the Wastewater Treatment Authority to dump on their property. The exact location is the facility off of Knapp and Old HW27 on the SW end of Houghton Lake.

6. Wild Rice Re-colonization:

One of the objectives in the current Houghton Lake management plan was to re-colonize the North Bay with a healthy, viable population of Wild Rice (*Zizania aquatica*). Previous presentations from Dr. Scott Herron from Ferris State University recommended that Muddy Bay would also be a favorable area for planting. RLS worked with Dr. Herron on the restoration of Wild Rice in Muddy Bay in 2020 evaluations in 2021 showed great success with germination. A follow-up survey in 2023 revealed that continued germination had declined, and this could be attributed to ice scouring and wind fetch due to the position in the lake basin and the very shallow depths. Surprisingly, Wild Rice colonization was naturally strong in the Middle Grounds in 2023-2024 and detailed surveys will be conducted again in 2025 to evaluate additional colonization and re-growth. There were approximately 40 acres of Wild Rice in the Middle Grounds located in 2024 which is a sign of strong natural colonization that may be possible due to lower water levels and also sustained control of the once-prevalent EWM in that region. The North Bay is also showing some natural growth and this area would be a suitable candidate for future seedings if sources are available. There are some additional areas of strong natural growth in the Muskegon River that were documented in 2024 and will also be evaluated. RLS is actively working on a scientific publication/peer-reviewed paper with Dr. Herron on this project as it contributes to lake restoration efforts and will share with the community and HLIB when completed.

Additional research on the lake sediments in 2023 revealed that sites where Wild Rice is present contained higher concentrations of ammonia nitrogen and phosphorus than uncolonized areas in the lake basin.

7. Boat Washing Stations:

RLS has recommended installation of boat washing stations at all points of entry to reduce the presence of invasive species into and out of Houghton Lake. Although this equipment is not patrolled regularly, it is of benefit if it is available for use. The HLIB and HLA are working together to determine the average use of each station and plan to promote increased use over time. This technology is an important tool for reducing herbicide treatment costs in the future.

8. Water Quality & Macroinvertebrate Monitoring:

Water quality parameters from the lake will also be monitored and graphed with historical data annually to observe long-term trends. In addition, water quality from the canals and tributaries will also be sampled. RLS will use that data to make any necessary recommendations for additional BMPs (best management practices) if needed.

The data collected to date have provided RLS and the HLIB with assurance that the lake is in overall good health. Sediment macroinvertebrates are good indicators of lake health and regular assessments allow for determination of lake health over time.

9. Educational Outreach for Houghton Lake:

RLS continues to assist the HLIB with an educational strategy to assist the Houghton Lake community with learning how to preserve and protect Houghton Lake. In 2019-2021, an educational ad campaign was released with the assistance of Spectrum® which was broadcast on local channels. RLS received feedback from many residents that the campaign was effective at raising awareness. RLS will continue to assist the HLIB with other educational opportunities with a community-wide workshop highly recommended in upcoming years. In 2025, a community-wide educational workshop will be held and discussed in the spring of 2025. The workshop will be held at a local venue and will include multiple informational booths, demonstrations, and provide updates on the success of the current Houghton Lake improvement program. RLS co-authored a cover feature article in the Michigan Riparian Fall, 2024 volume 60 (number 4) on Houghton Lake that summarized the historical and current aspects of the lake and its management and future direction.

10. PFAS in Houghton lake:

Recent concerns regarding the presence of PFAS in Houghton Lake have been addressed. In 2024, the MDNR tested fish tissue for PFAS, and it was found to be quite low in bluegill and perch. Areas of white foam have been observed in 2024 and should be monitored in future years. RLS has the ability to sample areas for PFAS given board approval. Figure 50 shows a dense foam of unknown origin on a canal in Houghton Lake in 2024. This foam could have been a surfactant from a storm drain but could also contain PFAS or other pollutants. Any discharge should be monitored in the future to protect the water quality of Houghton Lake.



Figure 50. A dense, white foam of unknown origin on a Houghton Lake canal (summer, 2024).

Table 23. Primary and Secondary Management Goals and Activities for each year of the 2022-2026 Houghton Lake Improvement Program.

Lake Management Activity	Primary Goal	Secondary Goal	Best Locations to Use
Aquatic herbicide treatment of hybrid milfoil	To reduce areas where the milfoil is prominent	To prevent dense areas from spreading in the lake	Main lake & canals
Aquatic Herbicide treatment of Starry Stonewort	To reduce areas where it is dense	To prevent plant from carpeting lake bottom	Main lake & canals
Mechanical Harvesting	Reduce dense areas in problem canals	Reduce DO depletion in canals	Canals
Benthic Barriers/Weed Rollers	To prevent germination of nuisance weeds in beach areas or canals	To reduce dependency on chemicals in nearshore areas	Beach areas, canals
Wild Rice Cultivation	To allow for new growth of Wild Rice in previously colonized areas	To increase biodiversity of native aquatic vegetation	Middle Grounds, North Bay, Muddy Bay
Phoslock® or SeClear® of canals	To reduce the presence of blue-green blooms in problem canals	To reduce nutrients that exacerbate blue-green blooms	Canals (especially MKP-5 canal system and Long Point canal as needed)
Lake Vegetation Surveys/Scans	To determine % cover by invasives and use as data tool for management	To compare year to year reductions in invasive vegetation areas	Main lake, canals
Boat Washing Stations	To clean boats of invasives before entering the lake	To educate boaters on the proper cleaning of boats and on invasives	All points of access as funding becomes available
Water Quality Lake & Tributary Monitoring	To troubleshoot areas that have poor water quality	To compare trend in water quality parameters with time	Main Lake, canals, tributaries
Macroinvertebrate Sampling	To determine changes in community structure as food source annually	To determine if herbicides have an impact on populations	Areas consistently sampled annually in main lake
Educational Outreach	To educate riparians and lake users on current lake health	To promote citizen lake protection	Proposed workshops in program years

Table 24. Proposed Houghton Lake improvement costs for the five year program.

<i>Proposed Houghton Lake Management Improvement Item</i>	<i>Estimated 2024-2026 Cost</i>
Herbicides for Hybrid Watermilfoil and Starry Stonewort and/or DASH Boat removal of invasives, Permit Fees	\$580,000
Professional Limnologist Services (limnologist surveys, sampling, contractor oversight, education)	\$75,000
Attorney Fees	\$2,500
Assessment Roll Mgmt.	\$4,000
Board Audit	\$3,400
Conferences	\$1,000
Insurance	\$2,600
Memberships	\$200
Printing/Publishing	\$4,000
Board Secretary	\$4,200
Board Treasurer	\$3,000
Office Supplies & Rent	\$2,100
Publications/Postage	\$3,250
Refunds	\$500
Travel Expenses	\$250
Boat Washing Support	\$20,000

TV/Radio	\$12,000
Wild Rice Restoration	\$10,000
Contingency (15%)	\$110,220
TOTAL ANNUAL ESTIMATED COST	\$837,200
APPROX. ANNUAL COST PER UNIT OF BENEFIT	\$183.27

Scientific Literature Cited

- Brunberg, A.K. 1995. Microbial activity and phosphorus dynamics in eutrophic lake sediments enriched with *Microcystis* colonies. *Freshwater Biology* 33: 541-555.
- Fallon, R.D., and T.D. Brock. 1981. Overwintering of *Microcystis* in Lake Mendota. *Freshwater Biology* 11:217-226.
- Higgins, S.N, Malkin, S.Y., Howell, E.T., Guildford, S.J., Campbell, L., Hiriart-Baer, V., and Hecky, R.E. 2008. An ecological review of *Cladophora glomerata* (Chlorophyta) in the Laurentian Great Lakes. *Journal of Phycology* 44(4): 839-854.
- Nizan, S., C. Dimentman, and M. Shilo. 1986. Acute toxic effects of the Cyanobacterium *Microcystis aeruginosa* on *Daphnia magna*. *Limnology and Oceanography* 31(3):497-502.
- Reynolds, C.S. 1984. *The Ecology of Freshwater Phytoplankton*. Cambridge University Press, Cambridge, UK.